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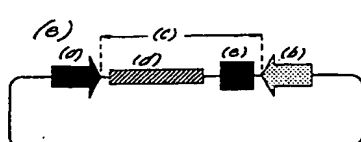
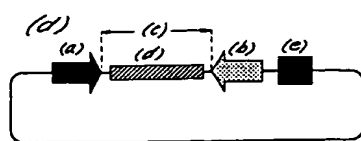
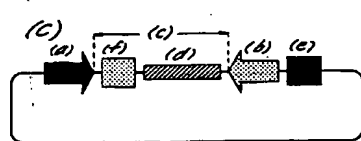
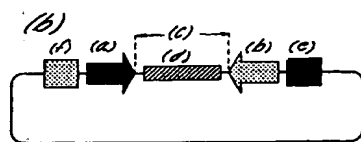
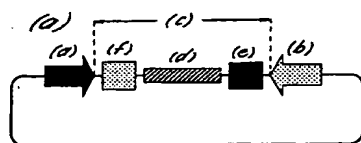
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(54) Title: **VECTOR CONSTRUCTS**



(57) Abstract: Vector constructs useful in the expression of double-stranded RNA. The constructs are particularly useful for expression of double-stranded RNA in vitro and in vivo.

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VECTOR CONSTRUCTS

Field of the invention

The invention relates to improved vector
5 constructs for use in the expression of double-
stranded RNA, particularly for use in the expression
of double-stranded RNA *in vitro* and *in vivo*.

Background to the invention

10 Since the advent of double-stranded RNA
inhibition (RNAi) as a tool for controlling gene
expression, as described in WO 99/32619 and WO
00/01846, there has been recognised a need for
specialised vectors designed for the production of
15 double-stranded RNA (dsRNA).

Cloning vectors designed to produce high levels
of dsRNA have been previously described by Plaetinck
et al. (WO 00/01846) and Timmons et al. Nature,
395:854 (1998). These vectors generally contain a
20 multiple cloning site (MCS) into which target DNA
fragments can be cloned flanked by two opposable
transcriptional promoters. Essentially, these three
components (Promoter 1, MCS and Promoter 2) make up
the entire system. In the appropriate expression
25 system, the DNA cloned into the MCS may be transcribed
in both directions, leading to the production of two
complementary RNA strands.

A disadvantage of the known systems is that not
only the cloned fragment is transcribed. Read-through
30 of the RNA polymerase will result in transcription of
the entire vector, and this also in both directions.
As only transcription of the cloned DNA fragment will
result in active dsRNA for RNAi purposes,
transcription of the vector part results in useless,

inefficient RNA. More specifically, 80% of these transcripts can be considered as non-specific and thus non-effective.

The large amounts of non-specific RNA generated by the prior art plasmid and expression systems results in some undesirable side effects. First, in RNAi protocols based on introduction of dsRNA into *C. elegans* via a food organism such as *E. coli* which expresses the dsRNA (see WO 00/01846), large RNA strands are considered to be toxic for the food organism. As a result, high amounts of RNA accumulating in *E. coli* cause a significant part of the population to die. Second, and probably more important, is the reduction of inhibition potential. The presence of large amounts of non-specific dsRNA causes a competitive environment for the specified sequences. The potential of the template-specified dsRNA sequences to inhibit the targeted protein expression in, for instance, *C. elegans* cells is reduced by the presence of these large non-specific regions. Such an inhibition by non-specific dsRNA has also been shown in *Drosophila* by Tushl et al., Genes & Development 13:3191-3197 (1999). Not only the potential to inhibit gene expression is affected, but also the amount of specific dsRNA produced is limited. Third, transcription of the vector backbone part, more particularly transcription of the origin of replication and related structures, results in plasmid instability and plasmid reorganisation, leading to reduced production of dsRNA. This relatively low concentration of effective dsRNA in turn leads to inefficient RNAi.

To conclude, the previously described vectors have following shortcomings: they are toxic to the

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feeding organism, a greater proportion of the transcripts produced are non-specific, the inhibitory potential of the dsRNA is reduced by the presence of non-specific regions, a high incidence of plasmid reorganizations and loss of plasmid from the feeding organism. It is therefore an object of the present invention to provide improved vectors for the production of dsRNA which avoid the disadvantages of the prior art vectors.

10 Vectors for use in the *in vitro* synthesis of RNA transcripts, for example the production of RNA probes, have been known and commonly used in the art for some time (see for example F. M. Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc. (1994); Jendrisak et al, Vectors for in vitro production of RNA copies of either strand of a cloned DNA sequence, US 4,766,072). In standard *in vitro* transcription protocols the problem of read-through transcription of vector sequences is generally avoided by linearizing the transcription vector at restriction site positioned at the 3' end of the desired transcript. However, this solution is not appropriate for *in vivo* transcription or for the production of dsRNA where it is important that the template is transcribed in both directions.

25 The inventors now propose a novel solution to the problems encountered with the prior art vectors for the production of dsRNA, based on the use of transcription terminators. Generally the solution consists of the use of at least one transcription terminator operably linked to at least one promoter, wherein the terminator stops the transcription initiated by the promoter. Any DNA fragment inserted between the 3' end of the promoter and the 5' end of

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the terminator will then be transcribed, without the unwanted transcription of the vector backbone.

Preferentially the vector consists of two promoters and two terminators, as further described below.

5 Therefore, in accordance with a first aspect of the invention there is provided a DNA construct comprising two opposable promoters flanking an inter-promoter region, the construct further comprising at least one transcription terminator positioned
10 transcriptionally downstream of one of the said promoters In particular, the invention provides for: a DNA construct comprising:

- a) a first promoter and
- b) a second promoter,

15 in which the first and second promoter are in opposite orientation to each other and define:

c) an inter-promoter region positioned downstream of the 3' end of the first promoter and downstream of the 3' end of the second promoter;

20 and which DNA construct further comprises:

d) at least one cloning site positioned in the inter-promoter region; and

e) a first transcription terminator, positioned (as seen from the 3' end of the first promoter)
25 downstream of the first promoter and downstream of the at least one cloning site, wherein the first transcription terminator is operably linked to the first promoter.

The inter-promoter region can also further be
30 defined as: the DNA region between the 3' end of the first promoter and the 3' end of the second promoter, and which is downstream of the first promoter, and which is downstream of the second promoter, and which preferably does not contains the 5' end of the first

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promoter and of the second promoter. The opposable first promoter and second promoter drive expression directional from their 5' ends to their 3' ends starting transcription downstream of their 3' ends, thus providing transcription of both strands of any nucleotide sequence(s) present in the inter-promoter region.

The two promoters present in the DNA construct of the invention may be identical or they may be different and may be of essentially any type. The precise nature of the promoters used in the construct may be dependent on the nature of the expression system in which the construct is expected to function (e.g. prokaryotic vs eukaryotic host cell). Bacteriophage promoters, for example the T7, T3 and SP6 promoters, are preferred for use in the constructs of the invention, since they provide advantages of high level transcription which is dependent only on binding of the appropriate RNA polymerase. Each of these promoters can independently be chosen. The phage promoters can also function in a wide variety of host systems, i.e. both prokaryotic and eukaryotic hosts, provided that the cognate polymerase is present in the host cell.

The arrangement of two "opposable" promoters flanking an inter-promoter region such that transcription initiation driven by one of the promoters results in transcription of the sense strand of the inter-promoter region and transcription initiation driven by the other promoter results in transcription of the antisense strand of the inter-promoter region is an arrangement well known in the art, for example, in the pGEM7 series of vectors from Promega Corp., Madison WI, USA.

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The DNA constructs of the invention differ from those of the prior art because of the presence of at least one transcription terminator positioned transcriptionally downstream of one of the promoters.

5 The transcription terminator may be uni- or bi-directional, the choice of uni- vs bi-directional terminators being influenced by the positioning of the terminator(s) within or outside the inter-promoter region, as explained below. The terminator may be of
10 prokaryotic, eukaryotic or phage origin. Bacteriophage terminators, for example T7, T3 and SP6 terminators, are particularly preferred. The only requirement is that the terminator must be capable of causing termination of transcription initiating at the
15 promoter relative to which it is transcriptionally downstream. In practice, these means that the promoter and terminator must form a 'functional combination', i.e. the terminator must be functional for the type of RNA polymerase initiating at the
20 promoter. By way of example, a eukaryotic RNA pol II promoter and a eukaryotic RNA pol II terminator would generally form a functional combination. The selection of a functional combination is particularly important where bacteriophage promoters and
25 terminators are to be used in the constructs of the invention, since the phage promoters and terminators are both polymerase-specific. To form a functional combination both the promoter and the terminator should be specific for the same polymerase, e.g. T7
30 promoter and T7 terminator, T3 promoter and T3 terminator etc.

In one embodiment, the DNA construct of the invention may comprise a single transcription terminator, positioned (as seen from the 3' end of the

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first promoter) downstream of the first promoter and downstream of the at least one cloning site, wherein the first transcription terminator is operably linked to the first promoter, wherein the single
5 transcription terminator is positioned in the inter-promoter region

In an alternative arrangement, the DNA construct comprises a single transcription terminator positioned outside of the inter-promoter region. In a still
10 further embodiment, the DNA construct may comprise two transcription terminators, each one of which is positioned transcriptionally downstream of one of the two promoters. In this arrangement, one or both of the terminators may be positioned within the inter-
15 promoter region. These various embodiments of the DNA constructs of the invention will be more fully described below, with reference to the accompanying drawings. The position of a first transcription terminator outside the inter-promoter region may also
20 be further defined as, i.e. such that a first transcription terminator is positioned (as seen from the 3' end of the first promoter) downstream of the first promoter, downstream of the at least one cloning site, and downstream of the 5' end of the second
25 promoter.

The position of a second transcription terminator outside the inter-promoter region may also be further defined as, i.e. such that a second transcription terminator positioned (as seen from the 3' end of the
30 second promoter) downstream of the second promoter, downstream of the at least one cloning site, and downstream of the 5' end of the first promoter.

Moreover, when the terminator is not located in the inter-promoter region, the distance between the 5'

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end of the first promoter and 3' end of the second terminator, or the distance between the 5' end of the second promoter and the 3' end of the first terminator is preferably small, i.e. such that the 3' end of the first transcription terminator is separated from the 5' end of the second promoter by no more than 2000 nucleotides, preferably no more than 1000 nucleotides, more preferably no more than 500 nucleotides, even more preferably no more than 200 nucleotides, especially preferably no more than 100 nucleotides, more especially preferable no more than 50 nucleotides, even more especially preferably no more than 20 nucleotides, particularly preferably no more than 10 nucleotides, more particularly preferably no more than 6 nucleotides.

Furthermore, when the second transcription terminator is located outside of the inter-promoter region, preferably the 3' end of the second transcription terminator is separated from the 5' end of the first promoter by no more than 2000 nucleotides, preferably no more than 1000 nucleotides, more preferably no more than 500 nucleotides, even more preferably no more than 200 nucleotides, especially preferably no more than 100 nucleotides, more especially preferably no more than 50 nucleotides, even more especially preferably no more than 20 nucleotides, particularly preferably no more than 10 nucleotides, more particularly preferably no more than 6 nucleotides

As defined above the term 'inter-promoter region' refers to all of the DNA sequence between the two promoters. As explained above, in certain embodiments of the invention the transcription terminator(s) may be sited within the inter-promoter region. The inter-

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promoter region may, advantageously, comprise a sequence of nucleotides forming a template for dsRNA production. The precise length and nature of this sequence is not material to the invention. The invention further provides DNA constructs in which the inter-promoter region comprises a cloning site. The function of the cloning site is to facilitate insertion of a DNA fragment forming a template for dsRNA production between the two promoters. Thus, the invention provides a series of cloning vectors which are of general use in the construction of template vectors for dsRNA production. Also encompassed within the scope of the invention are vectors derived from the cloning vectors which have a DNA fragment inserted into the cloning site.

The cloning site may further comprise one or more of the following:

- at least one restriction site, (as known in the art), or one or more further restriction sites, e.g. to provide a multiple cloning site (as known in the art),
- a stuffer DNA, e.g., flanked by at least two restriction site, such as two *Bst*XI restriction sites, or two *Xcm*I restriction sites,
- *att*R1 and *att*R2 recombination sites,
- a *ccdB* nucleotide sequence,
- a *ccdB* nucleotide further comprising at least one unique blunt-end restriction site, such as a *Srf*I restriction site, and/or

- a DNA fragment inserted in the at least one cloning site. All of the DNA constructs provided by the invention may, advantageously, form part of a replicable cloning vector, such as, for example, a plasmid vector. In addition to the opposable

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promoters, inter-promoter region and transcription terminator(s), the vector 'backbone' may further contain one or more of the general features commonly found in replicable vectors, for example an origin of replication to allow autonomous replication within a host cell and a selective marker, such as an antibiotic resistance gene. The selective marker gene (e.g. the antibiotic resistance gene) may itself contain a promoter and a transcription terminator and it is to be understood that these are completely independent of the promoter and terminator elements required by the invention and are not to be taken into consideration in determining whether a particular vector falls within the scope of the invention.

DNA constructs according to the invention may be easily be constructed from the component sequence elements using standard recombinant techniques well known in the art and described, for example, in F. M. Ausubel et al. (eds.), *Current Protocols in Molecular Biology*, John Wiley & Sons, Inc. (1994), as will be appreciated by one skilled in the art from the following detailed description of the invention and the accompanying Examples.

There follows a detailed description of DNA constructs according to the invention, with reference to the following schematic drawings in which:

Figures 1(a) to 1(e) are schematic representations of several different embodiments of the DNA construct according to the invention illustrating the relative positioning of the promoter and transcription terminator elements.

Figure 2(a) is a schematic representation of a prior

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art vector included for comparison purposes.

Figures 2(b) to 2(e) are schematic representations of several further embodiments of the DNA construct according to the invention illustrating the use of different cloning sites in the inter-promoter region.

Referring to the Drawings, Figure 1(a) schematically illustrates a first DNA construct according to the invention which is a plasmid vector comprising two opposable promoters; a first promoter a) and a second promoter b) flanking an inter-promoter region c), which inter-promoter region is downstream of the 3' of the first promoter, and downstream of the 3' end of the second promoter. The first promoter and the second promoter may be identical or different. This embodiment comprises a first transcription terminator e) and a second transcription terminator f) both of which are positioned within the inter-promoter region. In this embodiment, the first terminator and the second terminator are preferentially uni-directional terminators.

A DNA fragment may be inserted in the at least one cloning site d). Such fragment is subject to transcription directed by the first promoter a) and the second promoter b) (i.e. transcription of both strands), resulting in the generation of two RNA fragments which may combine to double-stranded RNA of the inserted DNA fragment (both *in vitro* and *in vivo*).

Any desired DNA sequence, such as a genomic DNA sequence, or a cDNA sequence or any other coding sequence, may be inserted in the at least one cloning site. Without being limited to any specific

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explanation, it is assumed that when a) and e) form a functional combination, RNA polymerase which initiates transcription at a) will transcribe the inter-promoter region including the at least cloning site and the DNA fragment inserted in the at least cloning site and will be terminated when it reaches e). Similarly, RNA polymerase which initiates transcription at b) will transcribe the inter-promoter region including the at least one cloning site and the DNA fragment inserted in the at least one cloning site and will terminate when it reaches f). The terminators cause the RNA polymerase to pause, stop transcription and fall off the template. This prevents the unlimited transcription of the vector backbone, and reduces the unspecific transcription of non-essential DNA.

The inter-promoter region further comprises a sequence of nucleotides corresponding to a target for double-stranded RNA inhibition. This sequence is designated 'TF' for target fragment. It is this sequence which, when transcribed into dsRNA, will be responsible for specific double-stranded RNA inhibition of a target gene. The target fragment may be formed from a fragment of genomic DNA or cDNA from the target gene. Its precise length and nucleotide sequence are not material to the invention.

In the arrangement shown in Figure 1(a) the two terminators are positioned on either side of the TF within the inter-promoter region. Each of the terminators is positioned transcriptionally downstream of one of the promoters, the first terminator e) is transcriptionally downstream of first promoter a) and the second terminator f) is transcriptionally downstream of the second promoter b). Assuming that a) and e) form a functional combination, as described

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above, RNA polymerase which initiates transcription at
a) will transcribe the inter-promoter region up to and
including TF and will be terminated when it reaches
e). Similarly, RNA polymerase which initiates
5 transcription at b) will transcribe the inter-promoter
region up to and including TF on the opposite strand
and will terminate when it reaches f). The
terminators cause the RNA polymerase to pause, stop
transcription and fall off the template. This
10 prevents the unlimited transcription of the vector
backbone, and reduces the unspecific transcription of
non-essential DNA.

The transcripts generated from this vector may,
depending on the precise placement of the terminators
15 in the vector, be almost completely specific dsRNAs
corresponding to the TF region. Through the direct
placement of the terminator sequences at the
downstream end of the TF region on both sides of the
inserted DNA fragment, the amount of material
20 transcribed is completely reduced to the
template-specified sequences. Therefore, a higher
amount of specific dsRNA is obtained. Furthermore the
constructs are now also more stable, due to the
non-transcription of the vector backbone. The latter
25 characteristic (stability), combined with the now
relatively higher specific transcription rate,
provides a system adapted to synthesise higher amounts
of specific short dsRNA strands. This proportionally
higher amount of transcript, resulting in high
30 concentrations of specific dsRNA, enhances the
inhibitory effect in RNAi protocols. In RNAi
protocols based on expression of dsRNA in a food
organism, toxicity for the feeding organisms due to
high RNA expression is brought to a minimal level by

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use of this vector.

A specific example of a vector of the type illustrated in Figure 1(a), considered by the inventors to be the optimal arrangement for RNAi applications, is plasmid pGN9 described in the accompanying Examples. The transcriptional terminators used in pGN9 are T7 RNA polymerase specific terminators, since the vector contains two opposable T7 promoters. However, other systems could be used such as an expression system based on the T3 or SP6 promoter, terminator and polymerase or other prokaryotic or eukaryotic promoters and terminators.

Figure 1(b) illustrates schematically a further DNA construct according to the invention which is a plasmid vector comprising two opposable promoters a) and b) flanking an inter-promoter region c). This vector also comprises two transcription terminators e) and f) but in this arrangement the two terminators are positioned outside of the inter-promoter region, in fact the terminator elements now flank the two promoters. The arrangement is such that e) is transcriptionally downstream of a) whilst f) is transcriptionally downstream of b). Here again e) terminates the transcription initiated by a), whilst f) terminates the transcription initiated by promoter b). Placement of the terminators outside of d) allows the use of bi-directional terminators as well as uni-directional terminators, in contrast to the arrangement in Figure 1(a) where uni-directional terminators are preferred because of the placement of the terminators between the promoters. A number of bi-directional terminators which could be used in accordance with the invention are known in the art.

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Generally these are observed to be polymerase non-specific.

The embodiment shown in Figure 1(b) provides essentially the same advantages as that shown in Figure 1(a) over the prior art vectors for dsRNA production. The vector shown in Figure 1(b) will lead to the production of transcripts which are slightly longer, including the promoter regions. This relatively small difference in the length of the transcript and hence the formed dsRNA will not severely affect the efficacy in an RNAi system.

The position of the terminators and promoter in the example as shown in figure 1 (b) are preferably placed at close proximity, such that the 5' end of the promoters are separated from the 3' end of the transcription terminators by no more than 2000 nucleotides, preferably no more than 1000 nucleotides, more preferably no more than 500 nucleotides, even more preferably no more than 200 nucleotides, especially preferably no more than 100 nucleotides, more especially preferably no more than 50 nucleotides, even more especially preferably no more than 20 nucleotides, particularly preferably no more than 10 nucleotides, more particularly preferably no more than 6 nucleotides.

Figure 1(c) illustrates schematically a further DNA construct according to the invention which is a plasmid vector comprising two opposable promoters a) and b) flanking an inter-promoter region c). In this embodiment one terminator (in this case e)) is positioned within the c) and the other (f)) is positioned outside c). Again, e) terminates transcription initiated by a) and f) terminates

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transcription initiated by b). This arrangement may provide a useful solution to the problem of one of the terminators interfering with polymerase activity in the other direction (e.g. f) interferes with b) initiated transcription). This vector essentially provides the same advantages as the vector variations shown in Figure 1(a) and Figure 1(b) over the prior art. The relatively small difference in the length of the transcript due to the transcription of one of the promoters will not significantly affect the efficacy in RNAi systems. This more particularly the case when the terminator that is located outside of the inter-promoter region c) is at close proximity of the promoter, as defined above.

Figures 1(d) and 1(e) illustrate schematically two further DNA constructs according to the invention which are both plasmid vectors comprising two opposable promoters a) and b) flanking an inter-promoter region c). These embodiments comprise a single terminator only. In the arrangement shown in Figure 1(d) a single terminator e) which terminates transcription from a) is placed outside of c). The placement of the terminator outside of the IPR allows the use of both a bi-directional terminator or a uni-directional terminator in this system. In the embodiment shown in Figure 1(d) e) is placed within the c). a) should therefore preferably be a uni-directional terminator.

Further embodiments of the DNA construct according to the invention are illustrated schematically in Figures 2(b) to 2(e).

These embodiments are all plasmid cloning vectors, based upon the optimal arrangement of promoters and terminators shown in Figure 1(a), and

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described above, containing cloning sites to facilitate the insertion of a DNA fragment into the at least on cloning site.

These embodiments are all plasmid cloning
5 vectors, based upon the optimal arrangement of promoters and terminators shown in Figure 1(a), containing cloning sites to facilitate the insertion of a target DNA fragment into the inter-promoter region.

10 Figure 2(a), which is a schematic representation of a prior art cloning vector, is included for comparison purposes. This vector comprises two opposable promoters a) and b), which may be identical or different, flanking a multi-cloning site (MCS).

15 Figure 2(b) illustrates a first type of plasmid cloning vector according to the invention. The vector contains a first opposable promoters a) and a second opposable promoter b) flanking an inter-promoter region. The inter-promoter region can further be
20 defined as: the DNA region between the 3' end of the first promoter and the 3' end of the second promoter, and which is downstream of the first promoter, and which is downstream of the second promoter, and which preferably does not contains the 5' end of the first
25 promoter and of the second promoter. The inter-promoter promoter region further comprises terminators e) and f) flanking a multi-cloning site MCS. The MCS comprises at least one individual restriction sites, an preferably more than one
30 restriction site as known in the art, any of which may be used for insertion of a DNA fragment.

Figure 2(c) illustrates a further type of plasmid cloning vector according to the invention. This vector again contains opposable promoters a) and b)

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flanking an inter-promoter region comprising terminators e) and f). In this embodiment, a) and b) flank a cloning site which is adapted for facilitated cloning of PCR fragments, comprising a stuffer DNA
5 flanked by two identical restriction sites, in this case BstXI sites. The specific sequence of the stuffer DNA is not essential, provided that said stuffer DNA does not interfere with the desired effect and/or the desired activity of the DNA constructs of the
10 invention. A specific example of a vector according to this aspect of the invention described herein is plasmid pGN29.

The cloning of PCR products using BstXI recognition sites and BstXI adaptors is generally
15 known in the art. The BstXI adaptors are commercially obtained, such as from Invitrogen (Groningen, the Netherlands). These adaptors are non-palindromic adapters designed for easier and efficient cloning of PCR products into vectors. These use of these adaptors
20 reduces the concatemerization of adapters or insert DNA by having non-complementary (CACA) overhangs. The stuffer DNA is included merely to allow easy differentiation between BstXI cut and uncut vector on the basis of size. Its precise length and sequence
25 are not of importance.

Figure 2(d) illustrates a further type of plasmid cloning vector according to the invention. This vector again contains opposable promoters a) and b) flanking an inter-promoter region comprising
30 terminators e) and f). In this embodiment, a) and b) flank a cloning site which facilitates "High Throughput" cloning based on homologous recombination rather than restriction enzyme digestion and ligation. As shown schematically in Figure 2(d), the cloning

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site comprises *attR1* and *attR2* recombination sites from bacteriophage lambda flanking a gene which is lethal to *E. coli*, in this case the *ccdB* gene.

5 An alternative cloning method of DNA fragments into this vector, (not shown in Figure 2 (d)), consists of a variant of this vector, wherein the *ccdB* DNA sequence further comprises at least one unique restriction site, preferably the at least unique restriction site is a blunt-end restriction site, such as a *SrfI* restriction site. Insertion of a DNA
10 fragment in the at least unique restriction, results in inactivation of the *ccdB* gene, and hence in inactivation of the lethal *ccdB* gene.

A further variant of a vector is shown in Figure
15 2(d) in which the *attR1* and the *attE2* are not present. Such a vector comprises an at least one cloning site, said at least one cloning site consisting of a *ccdB* sequence, said *ccdB* sequence further comprising at least one unique restriction site, preferably the at least unique restriction site is a blunt-end
20 restriction site, such as a *SrfI* restriction site. Insertion of a DNA fragment in the at least unique restriction, results in inactivation of the *ccdB* gene, and hence in inactivation of the lethal *ccdB* gene.

25 These cloning sites comprising the *ccdB* nucleotide sequence and/or the *attR* sites (*R1* and/or *R2*) are derived from the Gateway™ cloning system commercially available from Life Technologies, Inc. The Gateway™ cloning system has been extensively
30 described by Hartley et al. in WO 96/40724 (PCT/US96/10082). A specific example of a vector according to this aspect of the invention described herein is pGN39.

Figure 2(e) and 2(f) illustrate a still further type of plasmid cloning vector according to the invention. This vector again contains opposable promoters a) and b) flanking an inter-promoter region c) comprising terminators e) and f). In the embodiment shown in Figure 2(e), e) and f) flank a cloning site which facilitates "high throughput" cloning of PCR products by TATM cloning. This cloning site comprises a stuffer DNA flanked by two identical restriction sites for an enzyme which generates overhanging T nucleotides. In this case the restriction sites are XcmI sites, but other sites which are cleaved to generate overhanging T nucleotides could be used with equivalent effect. The overhanging T nucleotides facilitate cloning of PCR products which have a overhanging A nucleotide. This principle is known as TATM cloning. The cut vector with overhanging T nucleotides can be "topomerized" to generate a cloning vector of the type shown schematically in Figure 2(f), by linking the enzyme topoisomerase to the overhanging T nucleotides. The resulting vector also facilitates the cloning of PCR products by the principle known as TOPOTM cloning.

Both the TOPOTM and TATM cloning systems, although not for the vectors described in this invention, are commercially available from Invitrogen. The TOPOTM cloning system has extensively been described by Shuman in WO 96/19497 (PCT/US95/16099). The TATM cloning system has extensively been described by Hernstadt et al. in WO 92/06189 (PCT/US91/07147).

It will be readily appreciated by the skilled reader that whilst Figures 2(b)-2(f) illustrate the inclusion of different cloning sites into a vector of the type illustrated in Figure 1(a), these cloning

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sites could be included into any of the DNA constructs of the invention, including those illustrated schematically in Figures 1(b) to 1(e)

5 Application of the DNA constructs of the invention in RNAi technology.

As aforementioned, a major application of the DNA constructs/vectors of the invention is in the production of double stranded RNA for use in RNAi
10 technology. In particular, the constructs are useful in *in vivo* RNAi protocols in the nematode worm *C. elegans*.

In *C. elegans*, RNAi has traditionally been performed by injection dsRNA into the worm. Fire et
15 al. describes these methods extensively in International Application No. WO 99/32619. In short, both strands of RNA are produced *in vitro* using commercially available *in vitro* transcription kits. Both strands of RNA are allowed to form dsRNA, after
20 which the dsRNA is injected into *C. elegans*. The new vector system developed by the present inventors is a drastic improvement on this traditional method. First, the RNA can be produced in one step, for instance by using two identical promoters such as
25 in the vector pGN9. Second, and more important, due to the presence of terminators the transcripts, and hence the formed dsRNA, will be more specific as only the cloned target fragment will be transcribed. This will result in a more efficient RNAi.

30 A further method to perform RNAi experiments in *C. elegans* has been described by Plaetinck et al. in WO 00/01846. In this method *C. elegans* worms are fed with bacteria which produce dsRNA. The dsRNA passes the gut barrier of the worm and induces the same RNAi

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as if the dsRNA has been injected. For these experiments, the preferred *E. coli* strain is HT115 (DE3), and the preferred *C. elegans* strain is nuc-1;gun-1. The improved vectors provided by the invention also improve the efficiency of RNAi in this method, as shown in the example below, as only effective dsRNA is produced.

Another method to perform RNAi has also been described by Plaetinck et al. in WO 00/01846. In short, this method is based on the production of dsRNA in the worm itself. This can be done by using worm promoters in the described vectors, or by using a transgenic worm expressing a polymerase specific for non-*C. elegans* promoters present in the vector, such that this polymerase drives transcription of the dsRNA. The promoters will preferentially be selected from the known bacteriophage RNA promoters, such as T7 or T3 or SP6 RNA promoters, which provide the advantage of a high level of transcription dependent only on the binding of the cognate polymerase.

Plasmid vector DNA can be introduced into the worm by several methods. First, the DNA can be introduced by the traditional injection method (Methods in Cell Biology, Vol 48, *C. elegans* Modern Biological Analysis of an organism, ed. by Epstein and Shakes). Second, the DNA can be introduced by DNA feeding. As has been shown by Plaetinck et al. in WO 00/01846, plasmid DNA can be introduced into the worm by feeding the worm with an *E. coli* strain that harbors the plasmid. Preferentially the *E. coli* strain is OP50, or MC1061 or HT115 (DE3) but any other strain would suit for this purpose. The *C. elegans* strain is preferentially a nuc-1 mutant strain or a nuc-1; gun-1 strain. The plasmid DNA from the *E. coli*

passes the gut barrier and is introduced into the nematode, resulting in the expression of dsRNA. As with the other RNAi methods described above, the use of the new vector system will enhance the RNAi by the production of only specific dsRNA.

The invention will be further understood with reference to the following experimental Examples, together with the following additional Figures in which:

Figure 3 is a representation (plasmid map) of pGN1.

Figure 4 is a representation (plasmid map) of pGN9.

Figure 5 illustrates the nucleotide sequence of a fragment of plasmid pGN1, annotated to show the positions of the opposable T7 promoters.

Figure 6 depicts the nucleotide sequence of the T7 transcription terminator.

Figure 7 illustrates the sequences of oligonucleotides oGN27, oGN28, oGN29 and oGN30 used to insert T7 transcription terminators into pGN1. The positions of the T7 pol terminator sequences and of various restriction sites are marked.

Figure 8 illustrates the nucleotide sequence of a fragment of plasmid pGN9, annotated to show the positions of the opposable T7 promoters and the T7 transcription terminators.

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Figure 9 (a) is a representation (plasmid map) of pGN29; (b) is a representation (plasmid map) of pGN39; (c) is a representation (plasmid map) of the plasmid TopoRNAi.

5

Figure 10 shows the complete nucleotide sequence of plasmid pGN9.

10

Figure 11 shows the complete nucleotide sequence of plasmid pGN29.

Figure 12 shows the complete nucleotide sequence of plasmid pGN39.

15

Figure 13 shows the complete nucleotide sequence of plasmid TopoRNAi.

Figure 14 shows the complete sequence of plasmid pGN49A.

20

Figure 15 shows the complete sequence of plasmid pGN59A.

Figure 16 is a representation (plasmid map) of pGN49A.

25

Figure 17 is a representation (plasmid map) of pGN59A.

30

Example 1-Vector construction.

The starting point for construction of the
5 vectors exemplified herein was plasmid pGN1. This
plasmid, described in the applicant's co-pending
International Application No. WO 00/01846, contains
two opposable T7 promoters flanking a multi-cloning
site.

10 Vector construction was carried out according to
standard molecular biology techniques known in the art
and described, for example, in F. M. Ausubel et al.
(eds.), *Current Protocols in Molecular Biology*, John
Wiley & Sons, Inc. (1994).

1) Construction of pGN9

pGN1 was first digested with restriction enzymes
EcoRI and KpnI. Oligonucleotides oGN27 and oGN28
(Figure 7) were annealed to generate a double stranded
20 fragment which was then ligated into the EcoRI/KpnI
cut vector. The resulting plasmid was re-digested
with XbaI and HindIII. Oligonucleotides oGN29 and
oGN30 were annealed to generate a double-stranded
fragment which was then annealed into the XbaI/HindIII
25 cut vector. The resulting vector was designated pGN9
(Figures 4 and 10).

2) Construction of further cloning vectors

pGN29 (Figure 9(a); Figure 11) was generated by
30 replacing the MCS in pGN9 with a stuffer DNA flanked
by BstXI sites. BstXI adapters are commercially
available from Invitrogen (Groningen, the
Netherlands).

pGN39 (Figure 9 (b); Figure 12) generated by following steps; pGN29 was digested with BstXI. BstXI adapters (Invitrogen, Groningen, The Netherlands) were ligated to Cassette A provided by the GATEWAYTM system (Life Technologies, Inc.). Cassette A contains attR1, CmR, CcdA, CcdB, attR2. The Cassette A with the adapters were then ligated into the digested pGN29, resulting in pGN39A. pGN39A contains a unique SrfI site in the ccdB gene.

The TopoRNAi vector (figure 9 (c); figure 13) was generated in the following way; pGN29 was digested with BstXI. Using PCR with the primers oGN103 and oGN104 and template pCDM8 (Invitrogen, Groningen, The Netherlands), a stuffer was generated which includes XcmI sites. Onto the PCR product, BstXI adapters were ligated, and the resulting ligation product was ligated in the BstXI digested pGN29 vector resulting in the TopoRNAi vector.

oGN103: 5' TACCAAGGCTAGCATGGTTTATCACTGATAAGTTGG 3'

oGN104: 5' TACCAAGGCTAGCATGGGCCTGCCTGAAGGCTGC 3'

PGN49A was constructed to insert an additional unique non-blunt restriction site and to delete the CmR gene pGN39. Overlap PCR was used. A first PCR was performed with primers oGN126 and oGN127 and PGN39A as template. Using primers oGN128 and oGN129 and the same template a second fragment was generated. Overlap PCR using the resulting fragments and primers oGN126P and oGN129P resulted in a final PCR product. To this final PCR

- 27 -

product, BstXI adapters were ligated, and the ligation product was ligated into pGN29 digested with BstXI. The resulting vector was designated pGN49A.

- 5 A control vector was generated to test the efficiency of the pGN49A cloning vector, such vector should contain the T7 promoters, but not the T7 terminators. For this, the XbaI insert of pGN49A was isolated and cloned in pGN1 digested with the same restriction
10 enzyme. The resulting vector was designated pGN59A.

oGN126 pGATCTGGATCCGGCTTACTAAAAGCCAGATAACAGTATGC

oGN127 GGAGACTTTATCGCTTAAGAGACGTGCACTGGCCAGGGGGATCACC

oGN128:

- 15 CCAGTGCACGTCTCTTAAGCGATAAAGTCTCCCGTGAACCTTACCCGGTGG

oGN129 pGCTGTGTATAAGGGAGCCTGACATTTATATTCCCCAG

Example 2-To illustrate the usefulness of the improved vectors in RNA.

- 20 This experiment was designed to illustrate the improved efficiency of the improved vectors of this invention in double-stranded RNA inhibition, as compared to the vectors known from the prior art. A significant increase on the efficacy of the system
25 could be expected, as more effective dsRNA was produced and hence RNAi performed better. The experimental system for this proof of concept experiment was to measure *C. elegans* rescue at 25°C in nuc-1 / pha-1(e2123)ts *C. elegans* mutants by RNAi of
30 sup35 using dsRNA feeding of pGN-2 (-terminator) and pGN-12 (+ terminator), with PGN-1 (empty vector) as a control and dilutor. The pha-1 ts / sup-35 mutation has extensively been described by Schnabel in WO

99/49066.

The *nuc-1* mutation in *C. elegans* provides for a *C. elegans* strain exhibiting better uptake abilities, such as the uptake of the dsRNA complexes than wild type *C. elegans*. This mutant is deleted in the major DNase enzymes, and inventors have proven in earlier co-pending applications that this *C. elegans* strain results in enhanced RNAi by feeding the nematode with dsRNAs.

The *pha-1(e2123)ts* mutation provides a mutant *C. elegans* strain with a phenotype of survival at 15°C and lethality at 25°C. This lethality is suppressible by the inhibition of *sup-35* expression. RNAi of *sup-35* should thus facilitate the rescue of *pha-1(e2123)ts* at 25°C. The vectors of the present invention, when expressing dsRNA from *sup-35*, should increase the efficacy of *sup-35* RNAi in rescuing *pha-1(e2123)ts* mutants at 25°C, compared to vectors that do not contain the terminators.

Vector pGN1 was used as empty vector. Vector pGN2 (-terminator) is a vector harboring *sup-35* DNA and expressing *sup-35* dsRNA when introduced in the appropriate host, the vector has no transcriptional terminators inserted. Vector pGN12 (+ terminator) is a vector as described above, containing the transcriptional terminators, and hence resulting in improved dsRNA production when introduced into an appropriate host. Thus, this vector has two unidirectional transcriptional terminators, both placed inside of the inter-promoter region, and flanking the *sup-35* fragment. Use of the latter

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vector was expected to increase the efficacy of the system, here meaning a better rescue (survival) of pha-1(e2123)ts mutants at 25°C.

5 Experimental conditions

12-well micro-titer plates were filled with approximately 2 ml of NGM agar per well.

(1 liter of NGM agar: 15g Agar, 1g peptone, 3g NaCl, 1ml cholesterol solution (5 mg/ml in EtOH), with
10 sterile addition after autoclaving of 9.5 ml 0.1M CaCl₂, 9.5 ml 0.1 ml MgSO₄, 25 ml 1M KH₂PO₄/K₂HPO₄ buffer pH 6, Ampicillin (100 µg/l), 5ml 0,1M IPTG and 5 ml nystatin solution (dissolved 10 mg/ml in 1:1 EtOH:CH₃COONH₄ 7.5 M)

15

The dried plates were spotted with approximately 50 µl of an overnight culture of bacteria HT115 (DE3) (Fire A, Carnegie Institution, Baltimore, MD) transformed with the plasmids. Individual nematodes at the L4
20 growth stage were then placed in single wells at day 1. In each well 1 nematode (P1). At day two, the P1 nematodes were placed to a new well and left to incubate for a day. The same procedure was repeated at day 3. All plates were further incubated at 25°C to
25 allow offspring to be formed. Sup35 RNAi induced survival (rescue) was measured by counting the offspring.

Results

30 RNAi experiment in *C. elegans* nuc-1/pha-1(e2123)ts mutants by feeding with *E. coli* expressing sup-35 dsRNA.

- 30 -

Set up:

pGN1 as control

pGN2 (sup 35 - Term.)

pGN12 (sup 35 + Term.)

5

pGN2 + pGN1 dilutions 1/2, 1/4, 1/8, 1/16, 1/32

pGN12 + pGN1 dilutions 1/2, 1/4, 1/8, 1/16, 1/32

10 Conditions:

Incubation temperature 25°C

Readout:

Count offspring (adult hermaphrodites)

pGN1 (control)

Day 1	0	0	0	0
Day 2	0	0	0	0
Day 3	0	0	0	0

pGN2 (undiluted)

Day 1	12	4	48	32
Day 2	24	23	80	85
Day 3	5	0	9	16

pGN12 (undiluted)

Day 1	16	29	37	14
Day 2	27	22	57	2
Day 3	1	2	4	1

pGN 2+1, 1/2 dilution

Day 1	0	7	0	2
Day 2	9	10	0	3
Day 3	0	2	0	0

pGN 12+1, 1/2 dilution

Day 1	22	28	103	61
Day 2	36	45	53	40
Day 3	3	3	25	1

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pGN 2+1, 1/4 dilution

Day 1	28	23	0	0
Day 2	6	3	0	0
Day 3	0	0	0	0

pGN 12+1, 1/4 dilution

Day 1	*	6	36	5
Day 2		24	55	3
Day 3				

pGN 2+1, 1/8 dilution

Day 1	0	0	4	0
Day 2	0	0	11	0
Day 3	0	0	0	0

pGN 12+1, 1/8 dilution

Day 1	31	12	16	38
Day 2	4	5	37	4
Day 3	0	0	2	1

pGN 2+1, 1/16 dilution

Day 1	0	0	0	0
Day 2	0	0	0	1 little
Day 3	0	0	0	0

pGN 12+1, 1/16 dilution

Day 1	1	0	0	0
Day 2	2	0	0	1
Day 3	0	1	1	1

pGN 2+1, 1/32 dilution

Day 1	0	0	0	0
Day 2	0	0	0	0
Day 3	0	0	0	0

pGN 12+1, 1/32 dilution

Day 1	0	0	1	0
Day 2	0	L2	3	0
Day 3	2	0	L3- L4	0

5

* mother died

Conclusions

As expected, worms fed by bacteria harboring pGN1, did not result in the viable offspring, due to the lethal effect of the pha-1 mutation at this temperature.

- 5 Feeding the nematodes with *E. coli* harboring pGN2 or pGN12 both result in viable offspring. This is due to the feeding of the worm with dsRNA from sup-35. The remarkable difference between the two feeding experiments can be seen in the dilution series. When
10 diluting the bacteria harboring pGN2 with bacteria harboring pGN1, the number of offspring diminishes drastically, even at a low dilution of one to two. This dilution series indicates that high levels of dsRNA are needed to have a proper RNAi induction. In
15 the feeding experiment with bacteria harboring pGN12, significant offspring is still observed at a dilution of one to eight. This indicates that in the bacteria harboring pGN12, much more effective dsRNA is formed. This experiment clearly shows that the addition of
20 terminator sequences in vectors to express dsRNA as described above provide a significant advantage in the generation of RNAi.

25 Example 3: Comparison of RNAi efficiency of vectors with and without T7 terminators(pGN49 vs pGN59)

- Three different genes have been cloned in the vectors pGN49A and pGN59A. The cloning was performed by amplifying the gene fragments with PfuI DNA polymerase
30 producing blunt ends, facilitating cloning in these vectors. These PCR fragments were cloned into the vectors digested with SrfI. Correct fragment insertion of the clones was checked by PCR. The fragments are chosen such that ds expression and RNAi results in a

lethal phenotype of the offspring. This procedure allows to compare fast and easy the efficiency of the two vectors pGN49 and pGN59 in RNAi.

plasmid	Gene (acedb)	Vector backbone5
pGW5	B0511.8	pGN49A
pGW9	C01G8.7	pGN49A
pGW11	C47B2.3	pGN49A
pGW17	B0511.8	pGN59A
pGW21	C01G8.7	pGN59A
pGW23	C47B2.3	pGN59A

All the plasmids (pGW-series) are transformed in *E.coli* AB301-105 (DE3) bacteria by standard methodology. The bacteria are then grown in LB/amp at 37°C for 14-18h.
25 These cultures were centrifuged and the bacterial pellet dissolved in S-complete buffer containing 1mM IPTG and 100 µg/µl ampiciline.

In 96 well plates containing 100 µl S-complete
30 (containing 1 mM IPTG and 100 µg/µl ampiciline final concentration) and 10 µl of bacteria solution, 3 nematodes were added at each well, the nematodes were at the L1 growth stage.
The plates were incubated at 25°C for 5-6 days. Each

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day the plates are inspected for development of the larvae and the production of F1 offspring.

5 Results

The RNAi was performed in eight-fold for each constructed plasmid. The results show that when T7 terminators are inserted into the vector backbone, the expected phenotype gives a 100% occurrence. When T7 terminators are not used the reproducibility can decrease up to 50%. As in the previous experiment, the results show that the addition of terminators significantly enhances RNAi performance.

DNA

fragment	B0511.8	B0511.8	C01G8.7	C01G8.7	C47B2.3	C47B2.3
Vector	pGN49A	pGN59A	PGN49A	pGN59A	pGN49A	pGN59A
Resulting						
plasmid	PGW5	PGW17	PGW9	PGW21	PGW11	PGW23
Percentage						
lethal	100	75	100	87.5	100	50
Percentage						
offspring	0	25	0	12.5	0	50

PCR fragment generated by the primers oGN103 and
oGN104 on template pCDM8

TACCAAGGCT AGCATGGTTT ATCACTGATA AGTTGG
5 ATAAGTTGGT GGACATATTA TGTTTATCAG TGATAAAGTG TCAAGCATGA
CAAAGTTGCA GCCGAATACA GTGATCCGTG CCGGCCCTGG ACTGTTGAAC
GAGGTCGGCG TAGACGGTCT GACGACACGC AAAGTGGCGG AACGGTTGGG
GGTGCAGCAG CCGGCGCTTT ACTGGCACTT CAGGAACAAG CGGGCGCTGC
TCGACGCACT GGCCGAAGCC ATGCTGGCGG AGAATCATAC GCTTCGGTGC
10 CGAGAGCCGA CGACGACTGG CGCTCATTTT TGATCGGGAA TCCCGCAGCT
TCAGGCAGGC CCATGCTAGC CTTGGTACCA GCACAATGG

Overlap PCR Fragment, which was used to generate

15 pGN49A

gatctggatccggcttactaaaagccagataacagtatgcgtatttgcgcgctg
atTTTTgCGGTataagaatatatactgatatgtatacccgaaagtatgtcaaaaa
gagggtgtgctatgaagcagcgtattacagtgacagttgacagcgacagctatca
20 gttgctcaaggcatatatgatgtcaatatctcgggtctggttaagcacaaccatg
cagaatgaagcccgtcgtctgcgtgccgaacgctggaaagcggaaaatcaggaa
gggatggctgaggtcgcccgggtttattgaaatgaacggctcttttgctgacgag
aacagggactggtgaaatgcagtttaaggtttacacctataaaagagagagccg
ttatcgtctgtttgtggatgtacagagtgatattattgacacgcccgggcca
25 cggatggtgatccccctggccagtgcacgtctcttaagcgataaagtctccc
gtgaactttaccCGgtggtgcatatcggggatgaaagctggcgcatgatgac
caccgatatggccagtgtgccggtctccgttatcggggaagaagtggctgat
ctcagccaccgcgaaaatgacatcaaaaacgccattaacctgatgttctggg
gaatataaatgtcaggctcccttatacacagc

30

Claims:

1. A DNA construct comprising:
 - a) a first promoter and
 - 5 b) a second promoter,
in which the first and second promoter are in
opposite orientation to each other and define:
 - c) an inter-promoter region positioned downstream of
the 3' end of the first promoter and downstream of
10 the 3' end of the second promoter;
and which DNA construct further comprises:
 - d) at least one cloning site positioned in the inter-
promoter region; and
 - e) a first transcription terminator, positioned (as
15 seen from the 3' end of the first promoter)
downstream of the first promoter and downstream of
the at least one cloning site, wherein the first
transcription terminator is operably linked to the
first promoter.
- 20 2. A DNA construct according to claim 1, further
comprising:
 - f) a second transcription terminator positioned (as
seen from the 3' end of the second promoter)
25 downstream of the second promoter and downstream of
the at least one cloning site.
wherein the second transcription terminator is
operably linked to the second promoter.
- 30 3. A DNA construct according to claim 1 or 2, in
which the first transcription terminator is
positioned in the inter-promoter region.

4. A DNA construct according to claim 1 or 2, in which the first transcription terminator is positioned (as seen from the 3' end of the first promoter) downstream of the first promoter,
5 downstream of the at least one cloning site, and downstream of the 5' end of the second promoter.
5. A DNA construct according to any one of claims 2,
10 3 or 4, in which the second transcription terminator is positioned in the inter-promoter region.
6. A DNA construct according to any of claims 2, 3
15 or 4 in which the second transcription terminator is positioned (as seen from the 3' end of the second promoter) downstream of the second promoter, downstream of the at least one cloning site, and downstream of the 5' end of the first
20 promoter.
7. A DNA construct according to any one of claims 4,
5 or 6, in which the 3' end of the first transcription terminator is separated from the 5' end of the second promoter by no more than 2000
25 nucleotides, preferably no more than 1000 nucleotides, more preferably no more than 500 nucleotides, even more preferably no more than 200 nucleotides, especially preferably no more than 100 nucleotides, more especially preferably
30 no more than 50 nucleotides, even more especially preferably no more than 20 nucleotides, particularly preferably no more than 10

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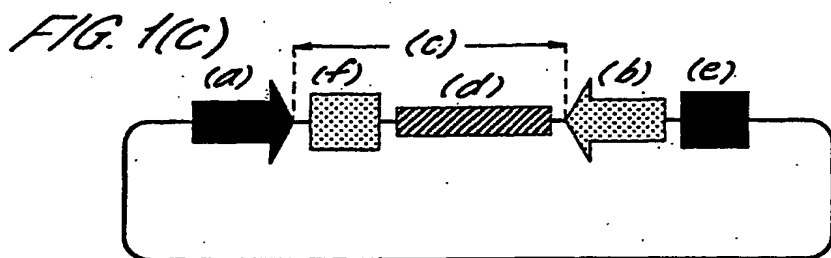
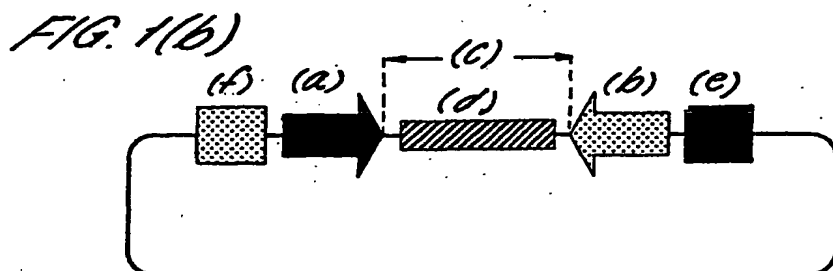
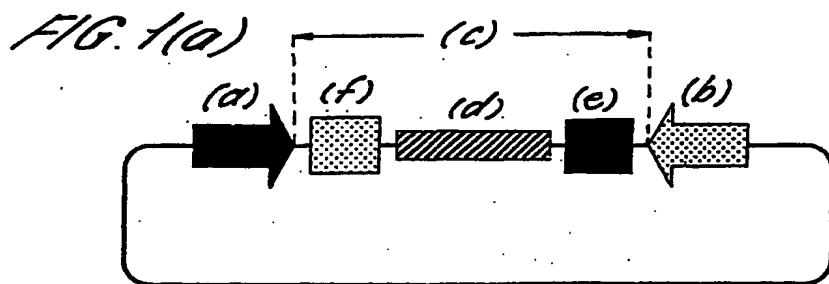
nucleotides, more particularly preferably no more than 6 nucleotides.

- 5 8. A DNA construct according to any one of claims 6 or 7, in which the 3' end of the second transcription terminator is separated from the 5' end of the first promoter by no more than 2000 nucleotides, preferably no more than 1000 nucleotides, more preferably no more than 500
10 nucleotides, even more preferably no more than 200 nucleotides, especially preferably no more than 100 nucleotides, more especially preferably no more than 50 nucleotides, even more especially preferably no more than 20 nucleotides,
15 particularly preferably no more than 10 nucleotides, more particularly preferably no more than 6 nucleotides.
- 20 9. A construct according to any one of the preceding claims wherein the first and the second promoter are identical.
- 25 10. A DNA construct according to any one of the claims 1 to 7 wherein the first and the second promoter are non-identical.
- 30 11. A DNA construct according to claims 8 or 9 wherein the first promoter and the second promoter are independently chosen from T7, T3 or SP6 promoters.
12. A construct according to any one of the preceding claims wherein the cloning site comprises at

least one restriction site.

13. A DNA according to claim 11 wherein the cloning
site comprises at least two restriction sites
5 flanking a sequence of stuffer DNA.
14. A DNA construct according to claim 12 wherein the
at least two restriction sites are identical.
- 10 15. A DNA construct according to any one of the claim
12 to 13 wherein the at least one restriction
site or the at least two restriction sites
restriction sites are *BstXI* sites.
- 15 16. A DNA construct according to any one of the
claims 12 to 13 wherein the restriction sites are
XcmI sites.
17. A DNA construct according to any one of the
20 preceding claims wherein the cloning site further
comprises *attR1* and *attR2* recombination
sequences.
18. A DNA construct according to any one of the
25 preceding claims wherein the cloning site further
comprises a *ccdB* nucleotide sequence.
19. A DNA construct according to claim 17 wherein the
ccdB nucleotide sequence further comprises at
30 least one unique restriction site.
20. A DNA construct according to claim 18 wherein the
at least one unique restriction site are blunt-
end restriction sites.

21. A DNA construct according to claim 19 wherein the blunt-end restriction sites are *SrfI* sites.
- 5 22. A DNA according to any one of the preceding claims which further comprises:
g) a DNA fragment inserted in the at least one cloning site.
- 10 23. A DNA construct according to any one of the preceding claims which is a plasmid or vector.
24. A plasmid or vector as claimed in claim 23 having the nucleotide sequence illustrated in Figure 10,
15 Figure 11, Figure 12, Figure 13, Figure 14, or Figure 15.
25. Use of the DNA construct according to any one of the preceding claims for the production of
20 double-stranded RNA for RNA inhibition.
26. A bacterial strain harbouring the DNA construct according to any one of the preceding claims.
- 25 27. A bacterial strain according to claim 26, wherein said bacteria strain is an *E. coli* strain.
28. Use of the bacterial strain according to claims 26 or 27 for the production of double-stranded
30 RNA for RNA inhibition.



(a): promoter 1
(b): promoter 2

(e): Terminator 1
(f): Terminator 2

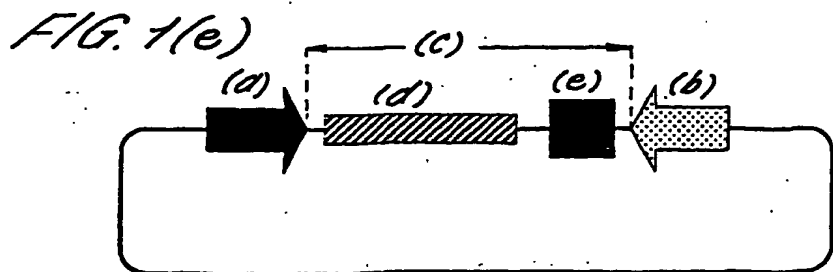
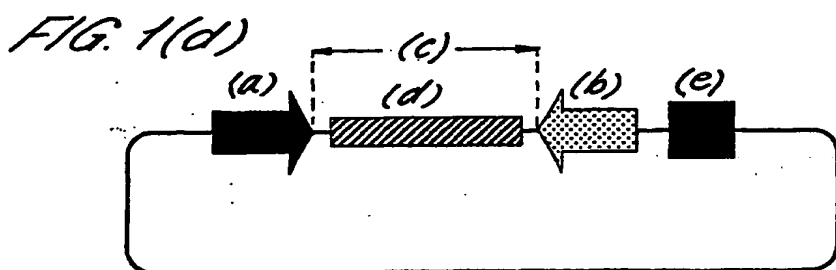


FIG. 2(a)

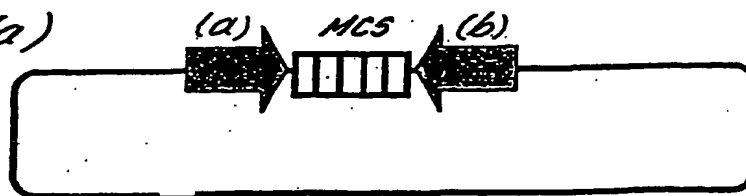


FIG. 2(b)

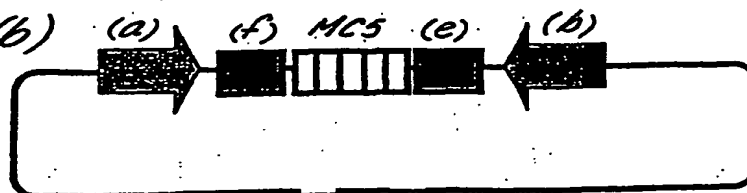


FIG. 2(c)

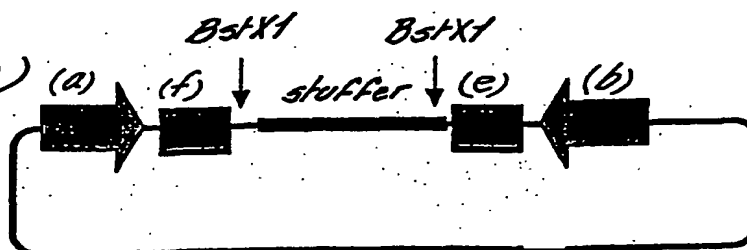


FIG. 2(d)

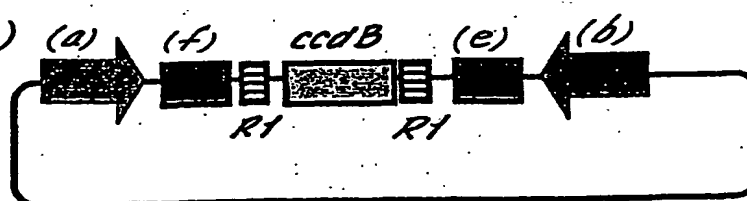


FIG. 2(e)

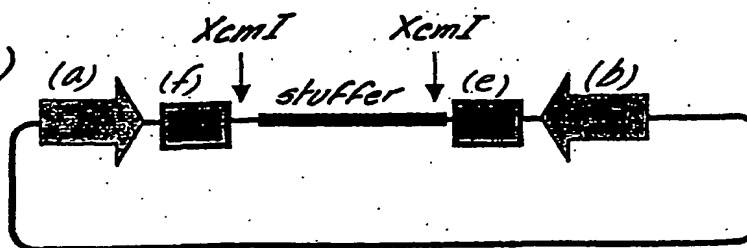
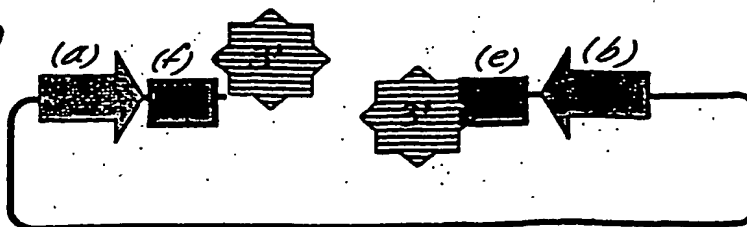


FIG. 2(f)



Construction RNAi vector with T7 terminators

FIG. 3.

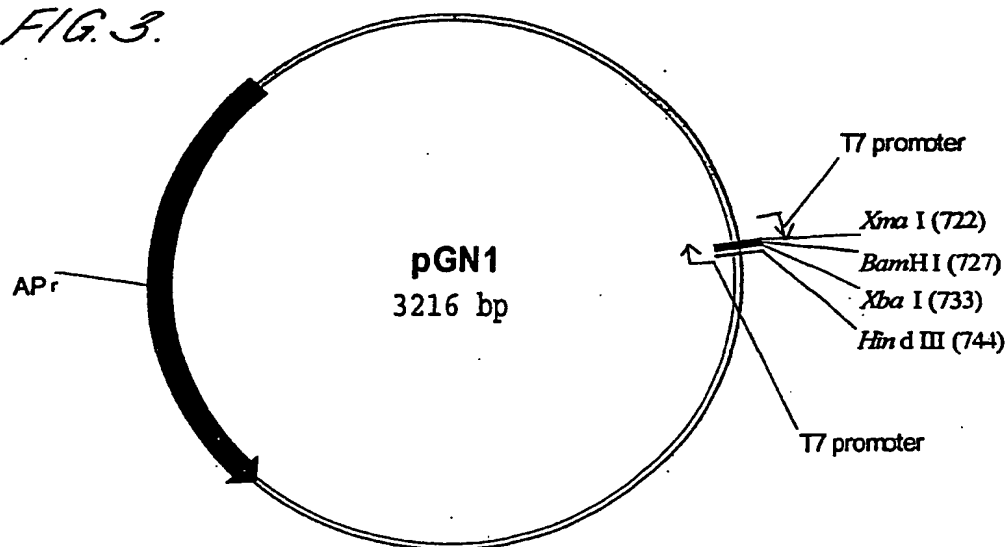


FIG. 4.

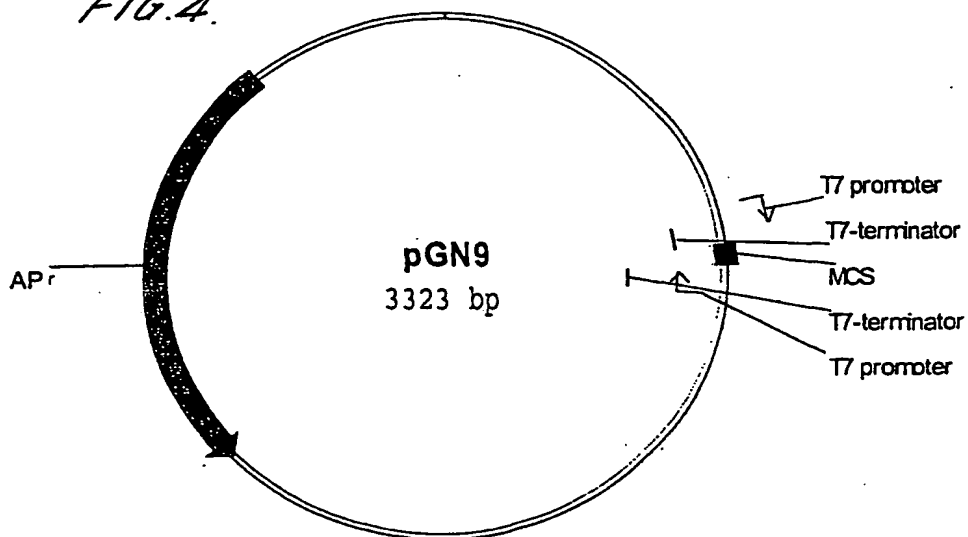


FIG. 5.

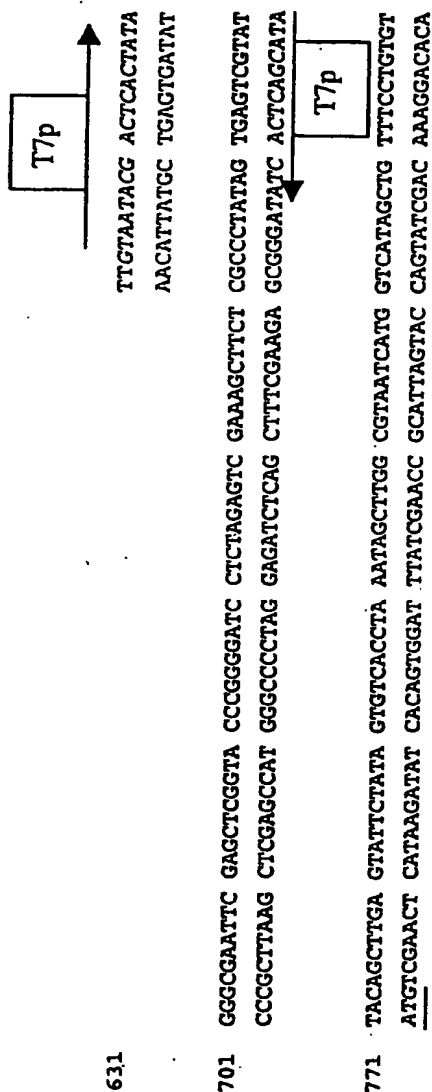
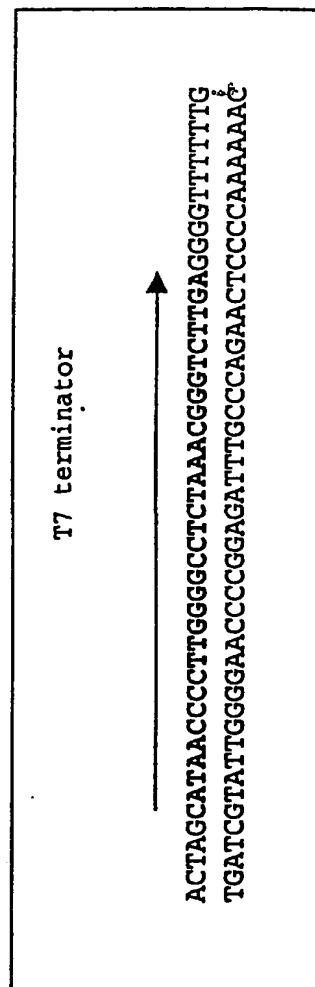
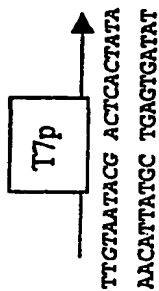


FIG. 6.



	EcoRI com.	T7 terminator	EcoRI PstI KpnI	
OGN27	5' AATCAAAAAACCCCTCAAGACCCCCTTAGAGGCCCAAGGGTTATGCTAGTGAATTCTGCAGCGGTAC			3'
OGN28	3' GTTTTTTGGGGAGTCTGGGCAAATCTCGGGGTTCCCCAATAACGATCACTTAAGACGTCGC			5'
		↓ T7 terminator ↑		
	XbaI MluI HindIII		HindIII com.	
OGN 29	5' CTAGACGCGTAAGCTTACTAGCATAACCCCTTGGGGCCCTCTAAACGGGTCCTTGAGGGGTTTTTG			3'
OGN 30	3' TGCGCATTCGAATGATCGTATTGGGGAAACCCCGGAGATTTGCCCCAGAACTCCCCAAAACCTCGA			5'

FIG. 8.



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701 GGGCGAATTC AAAAACCCTC TCAAGACCCG TTTAGAGGCC CCAAGGGGTT ATGCTAGTGA ATTCTGCAGG
CCCCGTTAAG TTTTCTGGG AGTCTGGG AAATCTCCGG GGTCCCCAA TACGATCACT TAAGAGGTCC



771 GTACCCGGG ATCCTCTAGA CGGTAAGCT TACTAGCATA ACCCTTGGG GCCTCTAAC GGTCTTGAG
CATGGGCCCC TAGGAGATCT GCGCATTCGA ATGATCGTAT TGGGGAACCC CGGAGATTG CCCAGAACTC



841 GGGTTTTTTG AGCTTCTCGC CCTATAGTGA GTGGTATTAC AGCTTGAGTA TTCTATAGTG TCACCTAAAT
CCCCAAAAAC TCGAAGAGCG GGATATCACT CAGCATATAG TCGAACTCAT AAGATATCAC AGTGGATTTA

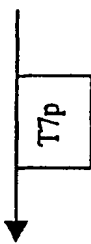


FIG. 9.

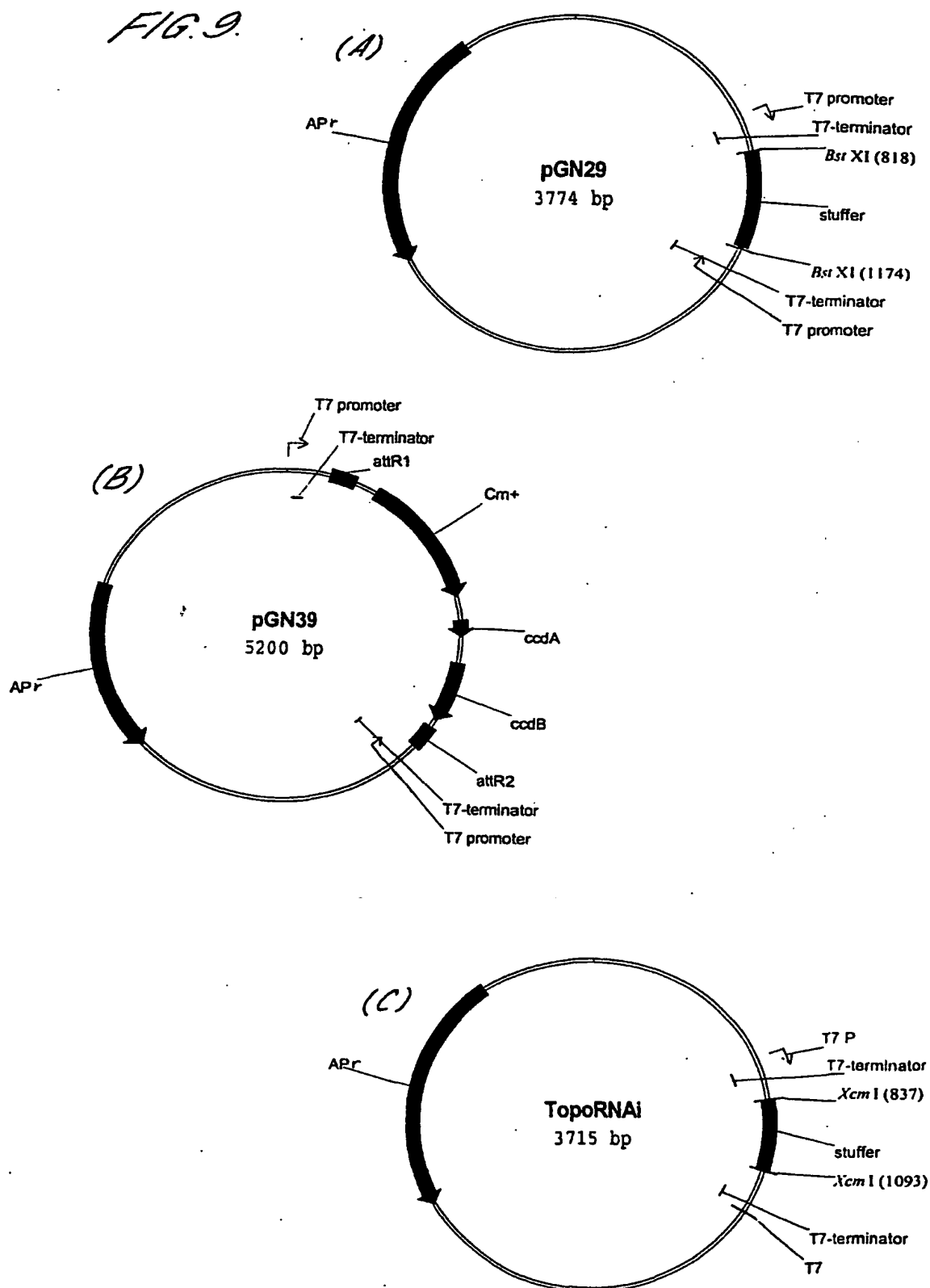


FIG. 10.

PGN9

1	gagtgcacca	tatgcggtgt	gaaataccgc	acagatgcgt	aaggagaaaa	taccgcatca
61	ggcgaaattg	taaacgttaa	tattttgtta	aaattcgcgt	taaatatttg	ttaaatacagc
121	tcaatttttta	accaataggc	cgaaatcggc	aaaatccctt	ataaatcaaa	agaatagacc
181	gagatagggt	tgagtgttgt	tccagtttgg	aacaagagtc	cactattaaa	gaacgtggac
241	tccaacgtca	aagggcgaaa	aaccgtctat	cagggcgatg	gcccactacg	tgaaccatca
301	cccaaatacaa	gttttttgcg	gtcagaggtgc	cgtaaagctc	taaatcggaa	ccctaaaggg
361	agcccccgat	ttagagcttg	acggggaag	ccggcgaaacg	tggcgagaaa	ggaagggaag
421	aaagcgaaaag	gagcgggcg	tagggcgctg	gcaagtgtag	cggtcacgct	gcgcgtaacc
481	accacacccg	ccgcgcttaa	tgcgcgccta	cagggcgctg	ccattcgcca	ttcaggctgc
541	gcaactgttg	ggaagggcga	tccggtcggg	cctcttcgct	attacgccag	ctggcgaaag
601	ggggatgtgc	tgcaaggcga	ttaagtgtgg	taacgccagg	gttttccag	tcacgacgtt
661	ggaaaacgac	ggccagtga	ttgtaatacg	actcactata	gggcgaattc	aaaaaacccc
721	tcaagacccg	tttagaggcc	ccaaggggtt	atgctagtga	attctgcagg	gtacccgggg
781	atcctctaga	cgcgtaagct	tactagcata	accccttggg	gcctctaaac	gggtcttgag
841	gggttttttg	agcttctcgc	cctatagtga	gtcgtattac	agcttgagta	ttctatagt
901	tcacctaaat	agcttggcgt	aatcatgggtc	atagctgttt	cctgtgtgaa	attgttatcc
961	gtcaccaatt	ccacacaaca	tacgagccgg	aagcataaag	tgtaaagcct	ggggtgccta
1021	atgagtgage	taactcacat	taattgcgtt	gcgctcactg	cccgccttcc	agtcgggaaa
1081	cctgtcgtgc	cagctgcatt	aatgaatcgg	ccaacgcgcg	gggagaggcg	gtttcgctat
1141	tgggcgctct	tccgcttcc	cgctcactga	ctcgtcgcgc	tccggtcggtc	ggctgcggcg
1201	agcgggtatca	gctcactcaa	aggcggtaat	acgggttatcc	acagaatcag	gggataacgc
1261	aggaagaaac	atgtgagcaa	aaggccagca	aaaggccagg	aaccgtaaaa	aggccgcgtt
1321	gctggcgctt	ttcgataggc	tccgcccccc	tgacgagcat	cacaaaaatc	gacgctcaag
1381	tcagagggtg	cgaaacccga	caggactata	aagataccag	gcgtttcccc	ctggaagctc
1441	cctcgtgcgc	tctcctgttc	cgaccctgcc	gcttaccgga	tacctgtccg	ccttctctcc
1501	ttcgggaagc	gtggcgcttt	ctcatagctc	acgctgtagg	tatctcagtt	cggtgtagg
1561	cggttcgctc	aagctgggct	gtgtgcacga	accccccggt	cagcccgacc	gctgcgcctt
1621	atccggtaac	tatcgtcttg	agtccaaccc	ggtaagacac	gacttatcgc	cactggcagc
1681	agccactggt	aacaggatta	gcagagcgag	gtatgtaggc	ggtgctacag	agttcttgaa
1741	gtgggtggcct	aactacggct	acactagaag	gacagtattt	ggtatctgcg	ctctgctgaa
1801	gccagttacc	ttcggaaaaa	gagttggtag	ctcttgatcc	ggcaaacaaa	ccaccgctgg
1861	tagcgggtgt	ttttttgttt	gcaagcagca	gattacgcgc	agaaaaaaag	gatctcaaga
1921	agatcctttg	atcttttcta	cggggtctga	cgctcagtg	aacgaaaact	cacgttaagg
1981	gatttttggtc	atgagattat	caaaaaggat	cttcacctag	atccttttaa	attaaaaatg
2041	aagtttttaa	tcaatctaaa	gtatatatga	gtaaaacttg	tctgacagtt	accaatgctt
2101	aatcagtgag	gcacctatct	cagcgatctg	tctatttcgt	tcatccatag	ttgcctgact
2161	ccccgtcgtg	tagataacta	cgatacggga	gggcttacca	tctggcccca	gtgctgcaat
2221	gataccgcga	gacccacgct	caccggctcc	agatttatca	gcaataaacc	agccagccgg
2281	aagggccgag	cgcagaagt	gtcctgcaac	tttatccgcc	tccatccagt	ctattaattg
2341	ttgccgggaa	gctagagtaa	gtagtccgcc	agttaatagt	ttgcgcaacg	ttgttggcat
2401	tgctacaggc	atcgtgggtg	cacgctcgtc	gtttggtagt	gcttcattca	gctccgggtc
2461	ccaacgatca	aggcgagtta	catgatcccc	catgttggtg	aaaaaagcgg	ttagctcctt
2521	cggtcctccg	atcgttgtca	gaagtaagtt	ggccgcagtg	ttatcactca	tggttatggc
2581	agcactgcat	aattctctta	ctgtcatgcc	atccgtaaga	tgcttttctg	tgactgggtga
2641	gtactcaacc	aagtcattct	gagaataccg	cgcccgcgga	ccgagttgct	cttgcccggc
2701	gtcaatacgg	gataatagt	tatgacatag	cagaacttta	aaagtgtca	tcattggaaa
2761	acgttctctg	gggcgaaaaa	tctcaaggat	cttaccgctg	ttgagatcca	gttcgatgta
2821	acccactcgt	gcacccaact	gatcttcagc	atcttttact	ttcaccagcg	tttctgggtg
2881	agcaaaaaaca	ggaaggcaaa	atgccgcaaa	aaagggaata	agggcgacac	ggaaatggtg
2941	aatactcata	ctcttccttt	ttcaatatta	ttgaagcatt	tatcagggtt	attgtctcat
3001	gagcggatac	atattttgaat	gtatttagaa	aaataaaca	ataggggttc	cgcgacatt
3061	tccccgaaaa	gtgccacctg	acgtctaaga	aaccattatt	atcatgacat	taacctataa
3121	aaatagcggt	atcacgagcg	cctttcgtct	cgcggttttc	gggtgatgacg	gtgaaaaacct
3181	ctgacacatg	cagctcccg	agacgggtcac	agcttgtctg	taagcggatg	ccgggagcag
3241	acaagcccg	cagggcgctg	cagcgggtgt	tggcgggtgt	cggggctggc	ttactatgc
3301	ggcatcagag	cagattgtac	tga			

FIG. 11.

PGN29

1	gagtgcacca	tatgcggtgt	gaaataccgc	acagatgcgt	aaggagaaaa	taccgcatca
61	ggcgaaattg	taaacgttaa	tatdddgtta	aaattcgcgt	taaatatttg	ttaaatcagc
121	tcatttttta	accaataggc	cgaaatcggc	aaaatccctt	ataaatcaaa	agaatagacc
181	gagatagggt	tgagtgttgt	tccagtttgg	aacaagagtc	cactattaaa	gaacgtggac
241	tccaacgtca	aagggcgaaa	aaccgtctat	cagggcgatg	gcccactacg	tgaaccatca
301	cccaaataca	gttttttgcg	gtcggaggtg	cgtaaagctc	taaatcggaa	ccctaagggg
361	agccccgat	ttagagcttg	acgggggaaag	ccggcgaaacg	tggcgagaaa	ggaagggaag
421	aaagcgaaa	gagcggggcg	tagggcgctg	gcaagtgtag	cgggtcacgct	gcgcgtaacc
481	accacacccg	ccgcgcctaa	tgccgcgcta	cagggcgcg	ccattcgcca	ttcaggctgc
541	gcaactgttg	ggaagggcga	tcgggtgcggg	cctcttcgct	attacgccag	ctggcgaaag
601	gggatgtg	tgcaaggcga	ttaaagttggg	taacgccagg	gttttcgag	tcacgacgtt
661	gtaaaacgac	ggccagtgaa	ttgtaatac	actcactata	gggcgaattc	aaaaaacccc
721	tcaagacccg	tttagaggcc	ecaaggggtt	atgctagtga	attctgcagg	gtaccggggg
781	atcctctaga	gatccctcga	cctcgagatc	cattgtgctg	gcgcggattc	tttatcactg
841	ataagttggt	ggacataatta	tgtttatcag	tgataaagt	tcaagcatga	caaagttgca
901	ccgaataaca	gtgatccgtg	ccggccctgg	actgttgaac	gaggtcggcg	tagacggtct
961	gacgacacgc	aaactggcgg	aacgggttggg	ggtgcagcag	ccggcgcttt	actggcactt
1021	caggaacaag	cgggcgctgc	tcgacgcact	ggccgaagcc	atgctggcgg	agaatcatac
1081	gcttcggtgc	cgagagccga	cgacgactgg	cgctcatttc	tgatcgggaa	tcccgcagct
1141	tcaggcaggc	gctgctcgcc	taccgcccag	acaatggatc	tcgagggatc	ttccatacct
1201	accagttctg	cgcttcgagg	tcgcccgcgc	gactctctag	acgcgtaagc	ttactagcat
1261	aaccctcttg	ggcctctaaa	cggtgtctga	ggggtttttt	gagcttctcg	ccctatagt
1321	agtctgatta	cagcttgagt	attctatagt	gtcacctaaa	tagcttggcg	taatcatggt
1381	catagctgtt	tcctgtgtga	aattgttatc	cgctcacaat	tccacacaac	atacgaagccg
1441	gaagcataaa	gtgtaaagcc	tggggtgcct	aatgagtgag	ctaactcaca	ttaattgcgt
1501	tgcgctcact	gcccgccttc	cagtcgggaa	acctgtcggt	ccagctgcag	taatgaatcg
1561	gccaacgcgc	ggggagaggg	gggttgcgta	ttggcgcttc	ttccgcttcc	tcgctcactg
1621	actcgctgcg	ctcggctcgt	cggtgcgggc	gagcggtatc	agctcactca	aaggcggtaa
1681	tacggttatc	cacagaatca	ggggataacg	caggaaagaa	catgtgagca	aaaggccagc
1741	aaaaggccag	gaaccgtaaa	aaggcccgct	tgctggcggt	tttcgatagg	ctccgcccc
1801	ctgacgagca	tcacaaaaat	cgacgctcaa	gtcagagggt	gcgaaacccg	acaggactat
1861	aaagatacca	ggcggtttccc	cctggaagct	ccctcggtcg	ctctcctggt	ccgacccctg
1921	cgcttacggg	atacctgtcc	gcctttctcc	cttcgggaag	cggtggcgctt	tctcatagct
1981	cacgctgtag	gtatctcagt	tcgggtgtagg	tcggtcgctc	caagctgggc	tgtgtgcaag
2041	aacccccctg	tcagcccgcg	cgctgcgcct	tatccggtaa	ctatcgctct	gagtccaacc
2101	cggttaagca	cgacttatcg	ccactggcag	cagccactgg	taacaggatt	agcagagcga
2161	ggtatgtagg	cggtgctaca	gagttcttga	agtgtgggcc	taactacggc	tacactagaa
2221	ggacagtatt	tggtatctgc	gctctgctga	agccagttac	cttcggaaaa	agagttggta
2281	gctcttgatc	cggaacacaa	accaccgctg	gtagcggtgg	tttttttgtt	tgcaagcagc
2341	agattacggc	cagaaaaaaa	ggatctcaag	aagatccttt	gatcttttct	acggggtctg
2401	acgctcagtg	gaacgaaaac	tcacgttaag	ggattttggt	catgagatta	tcaaaaagga
2461	tcctcaccta	gatcctttta	aattaaaaat	gaagttttta	atcaatctaa	agtatatatg
2521	agtaaaactg	gtctgacagt	taccaatgct	taatcagtg	ggcacctatc	tcagcgatct
2581	gtctatctcg	ttcatccata	gttgccctgac	tccccgctcg	gtagataaact	acgatacggg
2641	agggcttacc	atctggcccc	agtgtctgca	tgatacccg	agacccacgc	tcaccggctc
2701	cagattttatc	agcaataaac	cagccagccg	gaaggggcga	gcgcagaagt	ggtcctgcaa
2761	ctttatccgc	ctccatccag	tctattaatt	gttgccggga	agctagagta	agttagttcg
2821	cagttaatag	tttgcgcaac	gttggtggca	ttgctacagg	catcgtgggt	tcacgctcgt
2881	cgtttggtat	ggcttcattc	agctccggtt	cccaacgatc	aaggcgagtt	acatgatccc
2941	ccatgttggtg	caaaaaagcg	gttagctcct	tcggtcctcc	gatcgttggt	agaagtaagt
3001	tggccgcagt	gttatcactc	atgggttatgg	cagcactgca	taattctctt	actgtcatgc
3061	catccgtaag	atgcttttct	gtgactgggtg	agtactcaac	caagtcatte	tgagaatacc
3121	gcgcccggcg	accgagttgc	tcttgcccgtg	cgtcaatacg	ggataatagt	gtatgacata
3181	cgagaacttt	aaaagtgcct	atcatttgaa	aacgttcttc	ggggcgaaaa	ctctcagga
3241	tcctaccgct	gttgagatcc	agttcgatgt	aacccactcg	tgcacccaac	tgatcttcag
3301	catcttttac	tttcaccagc	gtttctgggt	gagcaaaaa	aggaaggcaa	aatgccgcaa
3361	aaaagggaat	aagggcgaca	cggaaatggt	gaatactcat	actcttctct	tttcaatatt
3421	attgaagcat	ttatcagggt	tattgtctca	tgagcggata	catatttgaa	tgtatttaga
3481	aaaataaaca	aataggggtt	ccgcgcacat	ttccccgaaa	agtgccacct	gacgtctaag
3541	aaaccattat	tatcatgaca	ttaaacctata	aaaataggcg	tatcacgagg	ccctttcgct
3601	tcgcgcggtt	cggtgatgac	ggtgaaaaac	tctgacacat	gcagctcccg	gcagcggtca
3661	cagcttgctc	gtaagcggat	gccgggagca	gacaagcccc	tcagggcgcg	tcagcggtg
3721	ttggcgggtg	tcggggctgg	cttaactatg	cggcatcaga	gcagattgta	ctga

*FIG. 12.**PGN39*

TAATACGACT CACTATAGGG CGAATTCAAA AAACCCCTCA AGACCCGTTT
AGAGGCCCCA AGGGGTTATG CTAGTGAATT CTGCAGCGGT ACCCGGGGAT
CCTCTAGAGA TCCCTCGACC TCGAGATCCA TTGTGCTGGA AAGATCACAA
GTTTGTACAA AAAAGCTGAA CGAGAAACGT AAAATGATAT AAATATCAAT
ATATTAAATT AGATTTTGCA TAAAAAACAG ACTACATAAT ACTGTAAAC
ACAACATATC CAGTCACTAT GCGCGCCGCA TTAGGCACCC CAGGCTTTAC
ACTTTATGCT TCCGGCTCGT ATAATGTGTG GATTTTGAGT TAGGATCCGG
CGAGATTTTC AGGAGCTAAG GAAGCTAAAA TGGAGAAAAA AATCACTGGA
TATACCACCG TTGATATATC CCAATGGCAT CGTAAAGAAC ATTTTGAGGC
ATTTTCAGTCA GTTGCTCAAT GTACCTATAA CCAGACCGTT CAGCTGGATA
TTACGGCCTT TTTAAAGACC GTAAAGAAAA ATAAGCACAA GTTTTATCCG
GCCTTTATTC ACATTCTTGC CCGCCTGATG AATGCTCATC CGGAATTCCG
TATGGCAATG AAAGACGGTG AGCTGGTGAT ATGGGATAGT GTTCACCCTT
GTTACACCGT TTTCCATGAG CAAACTGAAA CGTTTTTCATC GCTCTGGAGT
GAATACCACG ACGATTTCCG GCAGTTTCTA CACATATATT CGCAAGATGT
GGCGTGTTAC GGTGAAAACC TGGCCTATTT CCCTAAAGGG TTTATTGAGA
ATATGTTTTT CGTCTCAGCC AATCCCTGGG TGAGTTTCAC CAGTTTTGAT
TTAAACGTGG CCAATATGGA CAACTTCTTC GCCCCCGTTT TCACCATGGG
CAAATATTAT ACGCAAGGCG ACAAGGTGCT GATGCCGCTG GCGATTCAAG
TTCATCATGC CGTCTGTGAT GGCTTCCATG TCGGCAGAAT GCTTAATGAA
TTACAACAGT ACTGCGATGA GTGGCAGGGC GGGGCGTAAA GATCTGGATC
CGGCTTACTA AAAGCCAGAT AACAGTATGC GTATTTGCGC GCTGATTTTT
GCGGTATAAG AATATATACT GATATGTATA CCCGAAGTAT GTCAAAAAGA
GGTGTGCTAT GAAGCAGCGT ATTACAGTGA CAGTTGACAG CGACAGCTAT
CAGTTGCTCA AGGCATATAT GATGTCAATA TCTCCGGTCT GGTAAGCACA
ACCATGCAGA ATGAAGCCCG TCGTCTGCGT GCCGAACGCT GGAAAGCGGA
AATTACAGGA GGGATGGCTG AGGTCGCCCC GTTTATTGAA ATGAACGGCT
CTTTTGCTGA CGAGAACAGG GACTGGTGAA ATGCAGTTTA AGGTTTACAC
CTATAAAAGA GAGAGCCGTT ATCGTCTGTT TGTGGATGTA CAGAGTGATA
TTATTGACAC GCCCGGGCGA CGGATGGTGA TCCCCCTGGC CAGTGCACGT
CTGCTGTCAG ATAAAGTCTC CCGTGAACCT TACCCGCTGG TGCAATACGG
GGATGAAAGC TGGCGCATGA TGACCACCGA TATGGCCAGT GTGCCGGTCT
CCGTTATCGG GGAAGAAAGT GCTGATCTCA GCCACCGCGA AAATGACATC
AAAAACGCCA TTAACCTGAT GTTCTGGGGA ATATAAATGT CAGGCTCCCT
TATACACAGC CAGTCTGCAG GTCGACCATA GTGACTGGAT ATGTTGTGTT
TTACAGTATT ATGTAGTCTG TTTTTTATGC AAAATCTAAT TTAATATATT
GATATTTATA TCATTTTACG TTTCTCGTTC AGCTTTCTTG TACAAAGTGG
TGATCTTTCC AGCACAATGG ATCTCGAGGG ATCTTCATA CCTACCAGTT
CTGCGCCTGC AGGTCGCGGC CGCGACTCTA GACGCGTAAG CTTACTAGCA
TAACCCCTTG GGGCCTCTAA ACGGGTCTTG AGGGGTTTTT TGAGCTTCTC
GCCCTATAGT GAGTCGTATT ACAGCTTGAG TATTCTATAG TGTCACCTAA
ATAGCTTGGC GTAATCATGG TCATAGCTGT TTCCTGTGTG AAATGTTTAT
CCGCTCACAA TTCCACACAA CATACGAGCC GGAAGCATAA AGTGTAAGC

FIG. 12 (CONTINUED 1)

CTGGGGTGCC TAATGAGTGA GCTAACTCAC ATTAATTGCG TTGCGCTCAC
TGCCCGCTTT CCAGTCGGGA AACCTGTCGT GCCAGCTGCA TTAATGAATC
GGCCAAACGCG CGGGGAGAGG CGGTTTTCGT ATTGGGCGCT CTTCCGCTTC
CTCGCTCACT GACTCGCTGC GCTCGGTCGT TCGGCTGCGG CGAGCGGTAT
CAGCTCACTC AAAGGCGGTA ATACGGTTAT CCACAGAATC AGGGGATAAC
GCAGGAAAGA ACATGTGAGC AAAAGGCCAG CAAAAGGCCA GGAACCGTAA
AAAGGCCGCG TTGCTGGCGT TTTTCGATAG GCTCCGCCCC CCTGACGAGC
ATCACAAAAA TCGACGCTCA AGTCAGAGGT GGCGAAACCC GACAGGACTA
TAAAGATACC AGGCGTTTCC CCCTGGAAGC TCCCTCGTGC GCTCTCCTGT
TCCGACCCTG CCGCTTACCG GATACCTGTC CGCCTTTCTC CCTTCGGGAA
GCGTGGCGCT TTCTCATAGC TCACGCTGTA GGTATCTCAG TTCGGTGTAG
GTCGTTTCGT CCAAGCTGGG CTGTGTGCAC GAACCCCCCG TTCAGCCCCGA
CCGCTGCGCC TTATCCGGTA ACTATCGTCT TGAGTCCAAC CCGGTAAGAC
ACGACTTATC GCCACTGGCA GCAGCCACTG GTAACAGGAT TAGCAGAGCG
AGGTATGTAG GCGGTGCTAC AGAGTTCTTG AAGTGGTGGC CTAACACGG
CTACACTAGA AGGACAGTAT TTGGTATCTG CGCTCTGCTG AAGCCAGTTA
CCTTCGGAAA AAGAGTTGGT AGCTCTTGAT CCGGCAAACA AACCACCGCT
GGTAGCGGTG GTTTTTTTGT TTGCAAGCAG CAGATTACGC GCAGAAAAAA
AGGATCTCAA GAAGATCCTT TGATCTTTTC TACGGGGTCT GACGCTCAGT
GGAACGAAAA CTCACGTAA GGGATTTTGG TCATGAGATT ATCAAAAAGG
ATCTTCACCT AGATCCTTTT AAATTAAAAA TGAAGTTTTA AATCAATCTA
AAGTATATAT GAGTAACTT GGTCTGACAG TTACCAATGC TTAATCAGTG
AGGCACCTAT CTCAGCGATC TGTCTATTTC GTTCATCCAT AGTTGCCTGA
CTCCCCGTCTG TGTAGATAAC TACGATACGG GAGGGCTTAC CATCTGGCCC
CAGTGCTGCA ATGATACCGC GAGACCCACG CTCACCGGCT CCAGATTTAT
CAGCAATAAA CCAGCCAGCC GGAAGGGCCG AGCGCAGAAG TGGTCCTGCA
ACTTTATCCG CCTCCATCCA GTCTATTAAAT TGTGCGCGG AAGCTAGAGT
AAGTAGTTCG CCAGTTAATA GTTTGCGCAA CGTTGTTGGC ATTGCTACAG
GCATCGTGGT GTCACGCTCG TCGTTTGGA TGGCTTCATT CAGCTCCGGT
TCCCAACGAT CAAGGCGAGT TACATGATCC CCCATGTTGT GCAAAAAGC
GGTTAGCTCC TTCGGTCTCT CGATCGTTGT CAGAAGTAAG TTGGCCGCAG
TGTTATCACT CATGGTTATG GCAGCACTGC ATAATTCTCT TACTGTCATG
CCATCCGTAA GATGCTTTTC TGTGACTGGT GAGTACTCAA CCAAGTCATT
CTGAGAATAC CGCGCCCGGC GACCGAGTTG CTCTTGCCCG GCGTCAATAC
GGGATAATAG TGTATGACAT AGCAGAACTT TAAAAGTGCT CATCATTTGA
AAACGTTCTT CGGGGCGAAA ACTCTCAAGG ATCTTACCGC TGTGAGATC
CAGTTCGATG TAACCCACTC GTGCACCCAA CTGATCTTCA GCATCTTTTA
CTTTCACCAG CGTTTCTGGG TGAGCAAAAA CAGGAAGGCA AAATGCCGCA
AAAAAGGGAA TAAGGCGAC ACGGAAATGT TGAATACTCA TACTCTTCCT
TTTTCAATAT TATTGAAGCA TTTATCAGGG TTATTGTCCTC ATGAGCGGAT
ACATATTTGA ATGTATTTAG AAAAATAAAC AAATAGGGGT TCCGCGCACA
TTTCCCCGAA AAGTGCCACC TGACGTCTAA GAAACCATTA TTATCATGAC
ATTAACCTAT AAAAATAGGC GTATCACGAG GCCCTTTCGT CTCGCGCGTT
TCGGTGATGA CGGTGAAAAC CTCTGACACA TGCAGCTCCC GGAGACGGTC
ACAGCTTGTC TGTAAGCGGA TGCCGGGAGC AGACAAGCCC GTCAGGGCGC
GTCAGCGGGT GTTGGCGGGT GTCGGGGCTG GCTTAACTAT GCGGCATCAG

FIG. 12 (CONTINUED 2)

AGCAGATTGT ACTGAGAGTG CACCATATGC GGTGTGAAAT ACCGCACAGA
TGCCTAAGGA GAAAATACCG CATCAGGCGA AATTGTAAAC GTTAATATTT
TGTTAAAATT CGCGTTAAAT ATTTGTAAA TCAGCTCATT TTTTAACCAA
TAGGCCGAAA TCGGCAAAAT CCCTTATAAA TCAAAAGAAT AGACCGAGAT
AGGGTTGAGT GTTGTTCAG TTTGGAACAA GAGTCCACTA TTAAAGAACG
TGGACTCCAA CGTCAAAGGG CGAAAAACCG TCTATCAGGG CGATGGCCCA
CTACGTGAAC CATCACCCAA ATCAAGTTTT TTGCGGTCGA GGTGCCGTAA
AGCTCTAAAT CGGAACCCTA AAGGGAGCCC CCGATTTAGA GCTTGACGGG
GAAAGCCGGC GAACGTGGCG AGAAAGGAAG GGAAGAAAGC GAAAGGAGCG
GGCGCTAGGG CGCTGGCAAG TGTAGCGGTC ACGCTGCGCG TAACCACCAC
ACCCGCCGCG CTTAATGCGC CGCTACAGGG CGCGTCCATT CGCCATTGAG
GCTGCGCAAC TGTTGGGAAG GGCATCGGT GCGGGCCTCT TCGCTATTAC
GCCAGCTGGC GAAAGGGGGA TGTGCTGCAA GGCATTAAAG TTGGGTAACG
CCAGGGTTTT CCCAGTCACG ACGTTGTAAA ACGACGGCCA GTGAATTG

FIG. 13.

TopoRNAi

1	gagtgcacca	tatgcggtgt	gaaataccgc	acagatgcgt	aaggagaaaa	taccgcatca
61	ggcgaaattg	taaacggtta	tattttgtta	aaattcgcgt	taaatatttg	ttaaatacagc
121	tcatttttta	accaataggg	cgaaatcggc	aaaatccctt	ataaatcaaa	agaatagacc
181	gagataggg	tgagtgtgt	tccagtttgg	aacaagagtc	cactattaaa	gaacgtggac
241	tccaacgtca	aagggcgaaa	aaccgtctat	cagggcgatg	gcccactacg	tgaaccatca
301	cccaaatcaa	gttttttgcg	gtcgaaggtg	cgtaaagctc	taaatcgga	ccctaaaggg
361	agccccgat	ttagagcttg	acggggaaag	ccggcgaaag	tggcgagaaa	ggaagggaa
421	aaagcgaaa	gagcggggcg	tagggcgctg	gcaagtgtag	cggtcacgct	gcgcgttaac
481	accacacccg	ccgcgcttaa	tgccgcgcta	cagggcgcg	ccattcgcca	ttcaggctgc
541	gcaactgttg	ggaagggcga	tcgggtcggg	cctcttcgct	attacgccag	ctggcgaaag
601	ggggatgtgc	tgcaaggcga	ttaagttggg	taacgccagg	gttttcccag	tcacgacgtt
661	gtaaaacgac	ggccagtga	ttgtaatacg	actcactata	gggcgaattc	aaaaaacccc
721	tcaagacccg	tttagaggcc	ccaaggggtt	atgctagtga	attctgcagg	gtaccggggg
781	atcctctaga	gatccctcga	cctcgagatc	cattgtggtg	gaattctacc	aaggctagca
841	tgggcagccg	aatacagtga	tccgtgcggg	ccctggactg	ttgaacgagg	tcggcgtaga
901	cggtctgacg	acacgcaaac	tgccggaacg	gttgggggtg	cagcagccgg	cgctttactg
961	gcacttcagg	aacaagcggg	cgctgctcga	cgactggccc	gaagccatgc	tggcggagaa
1021	tcatacgctt	cggtgcccag	agccgacgac	gactggcgct	catttctgat	cgggaaatccc
1081	gcagccatgc	tagccttggt	agaattccac	cacaatggat	ctcgagggat	cttccatacc
1141	taccagttct	gcgcctgcag	gtcgcggccg	cgactctcta	gacgcgtaag	cttactagca
1201	taacccttg	gggcctctaa	acgggtcttg	aggggttttt	tgagcttctc	gccctatagt
1261	gagtcgtatt	acagcttgag	tattctatag	tgtaacctaa	atagcttggc	gtaatcatgg
1321	tcatagtgtg	ttcctgtgtg	aaattggtat	ccgctcacia	ttccacacaa	catacgagcc
1381	ggaagcataa	agtgtaaagc	ctggggtgcc	taatgagtga	gctaactcac	attaattgcg
1441	ttgcgctcac	tgcccgtttt	ccagtcggga	aacctgtcgt	gccagctgca	ttaatgaatc
1501	ggcgaacgcg	cggggagagg	cggtttgcgt	attgggcgct	cttcgccttc	ctcgctcact
1561	gactcgctgc	gctcggctcg	tcggtcgccg	cgagcgggtat	cagctcactc	aaagcgcgta
1621	atacggttat	ccacagaatc	aggggataac	gcaggaaaga	acatgtgagc	aaaagggccag
1681	caaaaggcca	ggaaccgtaa	aaaggcccg	ttgctggcgt	ttttcgatag	gctccgcccc
1741	cctgacgagc	atcacaaaaa	tcgacgctca	agtcagaggt	ggcgaaaccc	gacaggacta
1801	taaagatacc	aggcgtttcc	ccctggaagc	tccctcgctg	gctctcctgt	tccgaccctg
1861	ccgcttaccg	gatacctgtc	cgccctttct	ccttcgggaa	gcgtggcgct	ttctcatagc
1921	tcacgctgta	ggtatctcag	ttcgggttag	gtcgttcgct	ccaagctggg	cttctgtcac
1981	gaaccccccg	ttcagcccga	ccgctgcgcc	ttatccggta	actatcgtct	tgagtccaac
2041	ccggtaagac	acgacttatc	gccactggca	gcagccactg	gtaacaggat	tagcagagcg
2101	aggtatgtag	gcgggtgctac	agagttcttg	aagtgggtgg	ctaactacgg	ctacactaga
2161	aggcagtat	ttggatatctg	cgctctcttg	aagccagtta	acgttcgaaa	aagagttggg
2221	agctcttgat	ccggcaaaaca	aaccaccgct	ggtagcggtg	gtttttttgt	ttgcaagcag
2281	cagattacgc	gcagaaaaaa	aggatctcaa	gaagatcctt	tgatcttttc	tacgggggtc
2341	gacgctcagt	ggaacgaaaa	ctcacgttaa	gggattttgg	tcatgagatt	atcaaaaagg
2401	atcttcacct	agatcctttt	aaattaaaaa	tgaagtttta	aatcaatcta	aagtatatat
2461	gagtaaaact	ggtctgacag	ttaccaatgc	ttaatcagtg	aggcacctat	ctcagcgatc
2521	tgtctatttc	gttcacccat	agttgcctga	ctccccgtcg	tgtagataac	tacgatacgg
2581	gagggcctac	catctggccc	cagtgctgca	atgataccgc	gagaccacg	ctcaccgggt
2641	ccagatttat	cagcaataaa	ccagccagcc	ggaagggccg	agcgcagaag	tggtcctgca
2701	actttatccg	cctccatcca	gtctattaat	tggtgcccgg	aagctagagt	aagtagttcg
2761	ccagtttaata	gtttgcgcaa	cgttgttggc	attgctacag	gcacgtggt	gtcacgctcg
2821	tcgtttggtg	tggtttcatt	cagctccggg	tcccaacgat	caaggcgagt	tacatgatcc
2881	cccatgttgt	gcaaaaaagc	ggtagctccc	ttcgggtcctc	cgatcggtgt	cagaagtaag
2941	ttggccgcag	tggtatcact	catgggttat	gcagcactgc	ataattctct	tactgtcatg
3001	ccatccgtaa	gatgcttttc	tgtgactggt	gagtaactca	ccaagtcatt	ctgagaatac
3061	cgccgcccgc	gaccgagttg	ctcttgcccc	gcgtcaatac	gggataatag	tgatgatgat
3121	agcagaactt	taaaagtgtc	catcatttga	aaacgttctt	cggggcgaaa	actctcaagg
3181	atcttacccg	tggtgagatc	cagttcgatg	taaccacttc	gtgcacccaa	ctgatcttca
3241	gcatctttta	ctttcaccag	cgtttctggg	tgagcaaaaa	caggaaggca	aaatgccgca
3301	aaaaagggaa	taaggcgac	acggaaatgt	tgaataactca	tactcttctc	ttttcaatat
3361	tattgaagca	tttatcaggg	ttattgtctc	atgagcggat	acatatattga	atgtatttag
3421	aaaaataaac	aaataggggt	tcgcgcgaca	tttcccgaaa	aagtgccacc	tgacgtctaa
3481	gaaaccatta	ttatcatgac	attaacctat	aaaaataggc	gtatcacgag	gcccttctgt
3541	ctcgcgcggt	tcgggtgatga	cggtgaaaac	ctctgacaca	tgacgctccc	ggagacgggt
3601	acagcttgtc	tgtaagcgga	tcgcgggagc	agacaagccc	gtcagggcgc	gtcagcgggt
3661	gttggcgggt	gtcggggcgtg	gcttaactat	gcggcatcag	agcagattgt	actga

DGN49A *FIG. 14.*

TGTAATACGA CTCACTATAG GGCGAATTCA AAAAACCCTT CAAGACCCGT
TTAGAGGCCC CAAGGGGTTA TGCTAGTGAA TTCTGCAGCG GTACCCGGGG
ATCCTCTAGA GATCCCTCGA CCTCGAGATC CATTGTGCTG GAAAGGATCT
GGATCCGGCT TACTAAAAGC CAGATAACAG TATGCGTATT TCGCGCTGA
TTTTTTCGGT ATAAGAATAT ATACTGATAT GTATACCCGA AGTATGTCAA
AAAGAGGTGT GCTATGAAGC AGCGTATTAC AGTGACAGTT GACAGCGACA
GCTATCAGTT GCTCAAGGCA TATATGATGT CAATATCTCC GGTCTGGTAA
GCACAACCAT GCAGAATGAA GCCCGTCGTC TCGTGCCGA ACGCTGGAAA
GCGGAAAATC AGGAAGGGAT GGCTGAGGTC GCCCGGTTTA TTGAAATGAA
CGGCTCTTTT GCTGACGAGA ACAGGGACTG GTGAAATGCA GTTTAAGGTT
TACACCTATA AAAGAGAGAG CCGTTATCGT CTGTTTGTGG ATGTACAGAG
TGATATTATT GACACGCCCG GGCGACGGAT GGTGATCCCC CTGGCCAGTG
CACGTCTCTT AAGCGATAAA GTCTCCCGTG AACTTTACCC GGTGGTGCAT
ATCGGGGATG AAAGCTGGCG CATGATGACC ACCGATATGG CCAGTGTGCC
GGTCTCCGTT ATCGGGGAAG AAGTGGCTGA TCTCAGCCAC CGCGAAAATG
ACATCAAAAA CGCCATTAACT CTGATGTTCT GGGGAATATA AATGTCAGGC
TCCCTTATAC ACAGCCTTTC CAGCACAAATG GATCTCGAGG GATCTTCCAT
ACCTACCAGT TCTGCGCCTG CAGGTCGCGG CCGCGACTCT AGACGCGTAA
GCTTACTAGC ATAACCCCTT GGGGCTCTA AACGGGTCTT GAGGGGTTTT
TTGAGCTTCT CGCCCTATAG TGAGTCGTAT TACAGCTTGA GTATTCTATA
GTGTACCTA AATAGCTTGG CGTAATCATG GTCATAGCTG TTTCTGTGT
GAAATTGTTA TCCGCTCACA ATTCCACACA ACATACGAGC CGGAAGCATA
AAGTGTAAG CCTGGGGTGC CTAATGAGTG AGCTAACTCA CATTAAATTGC
GTTGCGCTCA CTGCCCGCTT TCCAGTCGGG AAACCTGTCT TGCCAGCTGC
ATTAATGAAT CGGCCAACGC GCGGGGAGAG GCGGTTTGCG TATTGGGCGC
TCTTCCGCTT CCTCGCTCAC TGAATCGCTG CGCTCGGTCT TTCGGCTGCG
GCGAGCGGTA TCAGCTCACT CAAAGGCGGT AATACGGTTA TCCACAGAAT
CAGGGGATAA CGCAGGAAAG AACATGTGAG CAAAAGGCCA GCAAAAGGCC
AGGAACCGTA AAAAGGCCGC GTTGCTGGCG TTTTTCGATA GGCTCCGCCC
CCCTGACGAG CATCACAAAA ATCGACGCTC AAGTCAGAGG TGGCGAAACC
CGACAGGACT ATAAAGATAC CAGGCGTTTC CCCCTGGAAG CTCCCTCGTG
CGCTCTCCTG TTCCGACCTT GCCGCTTACC GGATACCTGT CCGCCTTTCT
CCCTTCGGGA AGCGTGGCGC TTTCTCATAG CTCACGCTGT AGGTATCTCA
GTTGCGGTGA GGTGTTTCGC TCCAAGCTGG GCTGTGTGCA CGAACCCCCC
GTTGAGCCCG ACCGCTGCGC CTTATCCGGT AACTATCGTC TTGAGTCCAA
CCCGGTAAGA CACGACTTAT CGCCACTGGC AGCAGCCACT GGTAACAGGA
TTAGCAGAGC GAGGTATGTA GGCGGTGCTA CAGAGTTCTT GAAGTGGTGG
CCTAACTACG GCTACACTAG AAGGACAGTA TTTGGTATCT GCGCTCTGCT
GAAGCCAGTT ACCTTCGGAA AAAGAGTTGG TAGCTCTTGA TCCGGCAAAAC
AAACCACCGC TGGTAGCGGT GGTTTTTTTG TTTGCAAGCA GCAGATTACG
CGCAGAAAAA AAGGATCTCA AGAAGATCCT TTGATCTTTT CTACGGGGTC
TGACGCTCAG TGAACGAAA ACTCACGTTA AGGGATTTTG GTCATGAGAT
TATCAAAAAG GATCTTCACC TAGATCCTTT TAAATTAAAA ATGAAGTTTT

FIG. 14 (CONTINUED)

AAATCAATCT AAAGTATATA TGAGTAAACT TGGTCTGACA GTTACCAATG
CTTAATCAGT GAGGCACCTA TCTCAGCGAT CTGTCTATTT CGTTCATCCA
TAGTTGCCTG ACTCCCCGTC GTGTAGATAA CTACGATACG GGAGGGCTTA
CCATCTGGCC CCAGTGCTGC AATGATACCG CGAGACCCAC GCTCACCGGC
TCCAGATTTA TCAGCAATAA ACCAGCCAGC CGGAAGGGCC GAGCGCAGAA
GTGGTCCTGC AACTTTATCC GCCTCCATCC AGTCTATTAA TTGTTGCCGG
GAAGCTAGAG TAAGTAGTTC GCCAGTTAAT AGTTTGCGCA ACGTTGTTGG
CATTGCTACA GGCATCGTGG TGTACGCTC GTCGTTTGGT ATGGCTTCAT
TCAGCTCCGG TTCCCAACGA TCAAGGCGAG TTACATGATC CCCCATGTTG
TGCAAAAAAG CGGTTAGCTC CTTGGTCTCT CCGATCGTTG TCAGAAGTAA
GTTGGCCGCA GTGTTATCAC TCATGGTTAT GGCAGCACTG CATAATTCTC
TTACTGTCAT GCCATCCGTA AGATGCTTTT CTGTGACTGG TGAGTACTCA
ACCAAGTCAT TCTGAGAATA CCGCGCCCGG CGACCGAGTT GCTCTTGCCC
GGCGTCAATA CGGGATAATA GTGTATGACA TAGCAGAACT TAAAAAGTGC
TCATCATTGG AAAACGTTCT TCGGGGCGAA AACTCTCAAG GATCTTACCG
CTGTTGAGAT CCAGTTCGAT GTAACCCACT CGTGCACCCA ACTGATCTTC
AGCATCTTTT ACTTTCACCA GCGTTTCTGG GTGAGCAAAA ACAGGAAGGC
AAAATGCCGC AAAAAAGGGA ATAAGGGCGA CACGGAAATG TTGAATACTC
ATACTCTTCC TTTTTCATAA TTATTGAAGC ATTTATCAGG GTTATTGTCT
CATGAGCGGA TACATATTTG AATGTATTTA GAAAAATAAA CAAATAGGGG
TTCCGCGCAC ATTTCCCCGA AAAGTGCCAC CTGACGTCTA AGAAACCATT
ATTATCATGA CATTAAACCTA TAAAAATAGG CGTATCACGA GGCCCTTTCG
TCTCGCGCGT TCGGTGATG ACGGTGAAAA CCTCTGACAC ATGCAGCTCC
CGGAGACGGT CACAGCTTGT CTGTAAGCGG ATGCCGGGAG CAGACAAGCC
CGTCAGGGCG CGTCAGCGGG TGTGCGGGG TGTGCGGGCT GGCTTAACTA
TGCGGCATCA GAGCAGATTG TACTGAGAGT GCACCATATG CGGTGTGAAA
TACCGCACAG ATGCGTAAGG AGAAAATACC GCATCAGGCG AAATTGTAAA
CGTTAATATT TTGTTAAAAAT TCGCGTTAAA TATTTGTTAA ATCAGCTCAT
TTTTTAACCA ATAGGCCGAA ATCGGCAAAA TCCCTTATAA ATCAAAAGAA
TAGACCGAGA TAGGGTTGAG TGTTGTTCCA GTTTGGAACA AGAGTCCACT
ATTAAAGAAC GTGGAATCCA ACGTCAAAGG GCGAAAAACC GTCTATCAGG
GCGATGGCCC ACTACGTGAA CCATCACCCA AATCAAGTTT TTGCGGTCG
AGGTGCCGTA AAGCTCTAAA TCGGAACCCT AAAGGGAGCC CCCGATTTAG
AGCTTGACGG GGAAGCCGG CGAACGTGGC GAGAAAGGAA GGAAGAAAG
CGAAAGGAGC GGGCGCTAGG GCGCTGGCAA GTGTAGCGGT CACGCTGCGC
GTAACCACCA CACCCGCCGC GCTTAATGCG CCGCTACAGG GCGCGTCCAT
TCGCCATTCA GGCTGCGCAA CTGTTGGGAA GGGCGATCGG TGCGGGCCTC
TTTCGCTATTA CGCCAGCTGG CGAAAGGGGG ATGTGCTGCA AGGCGATTAA
GTTGGGTAAC GCCAGGGTTT TCCAGTCAC GACGTTGTAA AACGACGGCC
AGTGAAT

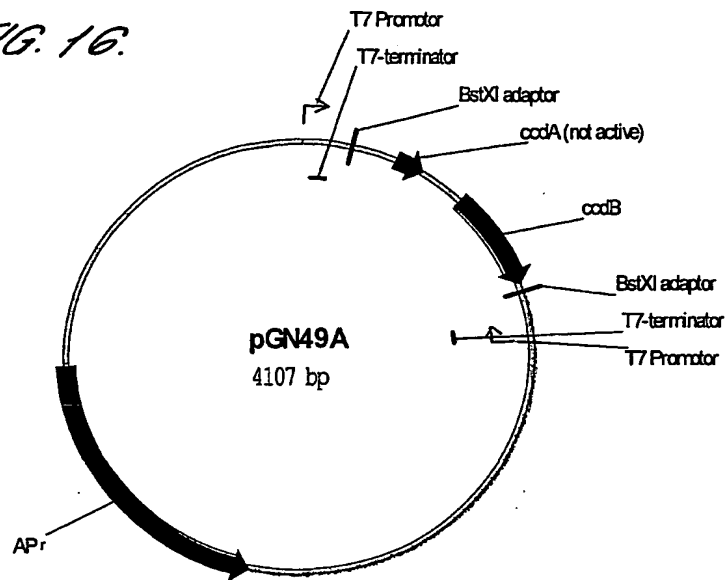
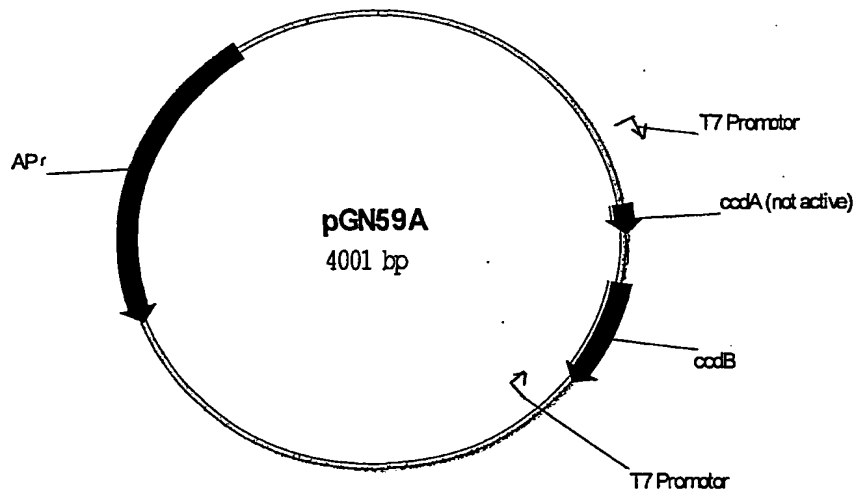
pgn59a *FIG. 15.*

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GAGTGCACCA TATGCGGTGT GAAATACCGC ACAGATGCGT AAGGAGAAAA
TACCGCATCA GGCGAAATTG TAAACGTTAA TATTTTGTTA AAATTCGCGT
TAAATATTTG TTAAATCAGC TCATTTTTTA ACCAATAGGC CGAAATCGGC
AAAATCCCTT ATAAATCAAA AGAATAGACC GAGATAGGGT TGAGTGTTGT
TCCAGTTTGG AACAAAGAGTC CACTATTAAA GAACGTGGAC TCCAACGTCA
AAGGGCGAAA AACCGTCTAT CAGGGCGATG GCCCACTACG TGAACCATCA
CCCAAATCAA GTTTTTTGCG GTCGAGGTGC CGTAAAGCTC TAAATCGGAA
CCCTAAAGGG AGCCCCGAT TTAGAGCTTG ACGGGGAAAG CCGGCGAACG
TGGCGAGAAA GGAAGGGAAG AAAGCGAAAG GAGCGGGCGC TAGGGCGCTG
GCAAGTGTAG CGGTACGCT GCGCGTAACC ACCACACCCG CCGCGCTTAA
TGCGCCGCTA CAGGGCGCGT CCATTGCGCA TTCAGGCTGC GCAACTGTTG
GGAAGGCGTA TCGGTGCGGG CCTCTCGCT ATTACGCCAG CTGGCGAAAG
GGGAGTGTGC TGCAAGGCGA TTAAGTTGGG TAACGCCAGG GTTTTCCAG
TCACGACGTT GTAAAACGAC GGCCAGTGAA TTGTAATACG ACTCACTATA
GGGCGAATTC GAGCTCGGTA CCCGGGGATC CTCTAGAGAT CCCTCGACCT
CGAGATCCAT TGTGCTGGAA AGGATCTGGA TCCGGCTTAC TAAAAGCCAG
ATAACAGTAT GCGTATTTGC GCGCTGATTT TTGCGGTATA AGAATATATA
CTGATATGTA TACCCGAAGT ATGTCAAAA GAGGTGTGCT ATGAAGCAGC
GTATTACAGT GACAGTTGAC AGCGACAGCT ATCAGTTGCT CAAGGCATAT
ATGATGTCAA TATCTCCGGT CTGGTAAGCA CAACCATGCA GAATGAAGCC
CGTCGTCTGC GTGCCGAACG CTGGAAGCG GAAAATCAGG AAGGGATGGC
TGAGGTCGCC CGGTTTATTG AAATGAACGG CTCTTTTGCT GACGAGAACA
GGGACTGGTG AAATGCAGTT TAAGGTTTAC ACCTATAAAA GAGAGAGCCG
TTATCGTCTG TTTGTGGATG TACAGAGTGA TATTATTGAC ACGCCCGGGC
GACGGATGGT GATCCCCCTG GCCAGTGCAC GTCTCTTAAG CGATAAAGTC
TCCCGTGAAC TTTACCCGGT GGTGCATATC GGGGATGAAA GCTGGCGCAT
GATGACCACC GATATGGCCA GTGTGCCGGT CTCCGTTATC GGGGAAGAAG
TGGCTGATCT CAGCCACCGC GAAAATGACA TCAAAAACGC CATTAACTG
ATGTTCTGGG GAATATAAAT GTCAGGCTCC CTTATACACA GCCTTTCCAG
CACAAATGGAT CTCGAGGGAT CTTCCATACC TACCAGTTCT GCGCCTGCAG
GTCGCGGCCG CGACTCTCTA GAGTCGAAAG CTTCTCGCCC TATAGTGAGT
CGTATTACAG CTTGAGTATT CTATAGTGTC ACCTAAATAG CTTGGCGTAA
TCATGGTCAT AGCTGTTTCC TGTGTGAAAT TGTTATCCGC TCACAATTCC
ACACAACATA CGAGCCGGAA GCATAAAGTG TAAAGCCTGG GGTGCCTAAT
GAGTGAGCTA ACTCACATTA ATTGCGTTGC GCTCACTGCC CGCTTTCCAG
TCGGGAAACC TGTCGTGCCA GCTGCATTAA TGAATCGGCC AACGCGCGGG
GAGAGGCGGT TTGCGTATTG GCGGCTCTTC CGCTTCCTCG CTCACTGACT
CGCTGCGCTC GGTGCTTCGG CTGCGGCGAG CGGTATCAGC TCACTCAAAG
GCGGTAATAC GGTTATCCAC AGAATCAGGG GATAACGCAG GAAAGAACAT
GTGAGCAAAA GGCCAGCAA AGGCCAGGAA CCGTAAAAAG GCCGCGTTGC
TGGCGTTTTT CGATAGGCTC CGCCCCCTG ACGAGCATCA CAAAATCGA
CGCTCAAGTC AGAGGTGGCG AAACCCGACA GGACTATAAA GATACCAGGC
GTTTCCCCCT GGAAGCTCCC TCGTGCGCTC TCCTGTTCCG ACCCTGCCGC
TTACCGGATA CCTGTCCGCC TTTCTCCCTT CGGGAAGCGT GCGGCTTTCT
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FIG. 15 (CONTINUED)

CATAGCTCAC GCTGTAGGTA TCTCAGTTCG GTGTAGGTCG TTCGCTCCAA
GCTGGGCTGT GTGCACGAAC CCCCCGTTCA GCCCGACCGC TGCGCCTTAT
CCGGTAACTA TCGTCTTGAG TCCAACCCGG TAAGACACGA CTTATCGCCA
CTGGCAGCAG CCACTGGTAA CAGGATTAGC AGAGCGAGGT ATGTAGGCGG
TGCTACAGAG TTCTTGAAGT GGTGGCCTAA CTACGGCTAC ACTAGAAGGA
CAGTATTTGG TATCTGCGCT CTGCTGAAGC CAGTTACCTT CGGAAAAAGA
GTTGGTAGCT CTTGATCCGG CAAACAAACC ACCGCTGGTA GCGGTGGTTT
TTTTGTTTGC AAGCAGCAGA TTACGCGCAG AAAAAAAGGA TCTCAAGAAG
ATCCTTTGAT CTTTTCTACG GGGTCTGACG CTCAGTGGAA CGAAACTCA
CGTTAAGGGA TTTTGGTCAT GAGATTATCA AAAAGGATCT TCACCTAGAT
CCTTTTAAAT TAAAAATGAA GTTTTAAATC AATCTAAAGT ATATATGAGT
AAACTTGGTC TGACAGTTAC CAATGCTTAA TCAGTGAGGC ACCTATCTCA
GCGATCTGTC TATTTCTGTC ATCCATAGTT GCCTGACTCC CCGTCGTGTA
GATAACTACG ATACGGGAGG GCTTACCATC TGGCCCCAGT GCTGCAATGA
TACCGCGAGA CCCACGCTCA CCGGCTCCAG ATTTATCAGC AATAAACAG
CCAGCCGAA GGGCCGAGCG CAGAAGTGGT CCTGCAACTT TATCCGCCTC
CATCCAGTCT ATTAATGTGT GCCGGGAAGC TAGAGTAAAGT AGTTCGCCAG
TTAATAGTTT GCGCAACGTT GTTGGCATTG CTACAGGCAT CGTGGTGTC
CGCTCGTCGT TTGGTATGGC TTCATTGAGC TCCGGTTCCC AACGATCAAG
GCGAGTTACA TGATCCCCCA TGTGTGCAA AAAAGCGGTT AGCTCCTTCG
GTCCTCCGAT CGTTGTCAGA AGTAAGTTGG CCGCAGTGTT ATCACTCATG
GTTATGGCAG CACTGCATAA TTCTCTTACT GTCATGCCAT CCGTAAGATG
CTTTTCTGTG ACTGGTGAGT ACTCAACCAA GTCATTCTGA GAATACCGCG
CCCGGCGACC GAGTTGCTCT TGCCCGGCGT CAATACGGGA TAATAGTGTA
TGACATAGCA GAACTTTAAA AGTGCTCATC ATTGGAAAAC GTTCTTCGGG
GCGAAAACCTC TCAAGGATCT TACCGCTGTT GAGATCCAGT TCGATGTAAC
CCACTCGTGC ACCCAACTGA TCTTCAGCAT CTTTTACTTT CACCAGCGTT
TCTGGGTGAG CAAAAACAGG AAGGCAAAAT GCCGCAAAA AGGGAATAAG
GGCGACACGG AAATGTTGAA TACTCATACT CTTCTTTTTT CAATATTATT
GAAGCATTTA TCAGGGTTAT TGTCTCATGA GCGGATACAT ATTTGAATGT
ATTTAGAAAA ATAAACAAAT AGGGGTTCCG CGCACATTC CCCGAAAAGT
GCCACCTGAC GTCTAAGAAA CCATTATTAT CATGACATTA ACCTATAAAA
ATAGGCGTAT CACGAGGCC TTTCTGCTCG CGCGTTTCGG TGATGACGGT
GAAAACCTCT GACACATGCA GCTCCCGAG ACGGTCACAG CTTGTCTGTA
AGCGGATGCC GGGAGCAGAC AAGCCCGTCA GGGCGCGTCA GCGGGTGTG
GCGGGTGTCG GGGCTGGCTT AACTATGCGG CATCAGAGCA GATTGTACTG

A

FIG. 16.*FIG. 17.*

SEQUENCE LISTING

<110> DEVGEN NV

<120> VECTOR CONSTRUCTS

<130> SCB/55178/001

<140>

<141>

<160> 21

<170> PatentIn Ver. 2.0

<210> 1

<211> 160

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Fragment of
pGN1 containing opposable T7 promoters

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gaaagcttct cgccctatag tgagtcgtat tacagcttga gtattctata gtgtcaccta 120
aatagcttgg cgtaatcatg gtcatactgt tttcctgtgt 160

<210> 2

<211> 49

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: DNA sequence
containing a T7 terminator

<400> 2

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<210> 3

<211> 70

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide oGN27

<400> 3

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gcagcggtag 70

<210> 4

<211> 62

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:

Oligonucleotide oGN28

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tg 62

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<211> 65
<212> DNA
<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide oGN29

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ttttg 65

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<211> 65
<212> DNA
<213> Artificial Sequence

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<223> Description of Artificial Sequence:
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cgcgt 65

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<211> 230
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Fragment of
plasmid pGN9 containing opposable T7 promoters and
T7 transcription terminators

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tactagcata accccttggg gcctctaaac gggctctgag ggggtttttg agcttctcgc 180
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<210> 8
<211> 3323
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence: Plasmid pGN9

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<211> 3774

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Plasmid pGN29

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<211> 5148

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Plasmid pGN39

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<211> 3715

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Plasmid
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<400> 11

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<211> 4107

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Plasmid pGN49A

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<213> Artificial Sequence

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<223> Description of Artificial Sequence:
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<223> Description of Artificial Sequence:
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<223> Description of Artificial Sequence: PCR fragment
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ccctggactg ttgaacgagg tcggcgtaga cggctctgacg acacgcaaac tggcggaacg 180
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<212> DNA
<213> Artificial Sequence

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<223> Description of Artificial Sequence: PCR fragment

<400> 21

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tatggccagt	gtgccggtct	ccgttatcgg	ggaagaagtg	gctgatctca	gccaccgcga	600
aaatgacatc	aaaaacgcca	ttaacctgat	gttctgggga	atataaatgt	caggctccct	660
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INTERNATIONAL SEARCH REPORT

International Application No

PCT/IB 01/01068

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 C12N15/10 C12N15/63 C12N15/70 C12N1/21

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the International search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS, CHEM ABS Data, MEDLINE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FR 2 782 325 A (PROTEUS) 18 February 2000 (2000-02-18) page 7, line 20 -page 8, line 8 page 11, line 11 - line 36 page 23, line 31 -page 24, line 9 ---	1-24, 26, 27
A	WO 00 01846 A (DEVGEN N.V.) 13 January 2000 (2000-01-13) cited in the application page 8, line 9 -page 10, line 22 page 15, line 9 - line 33 page 21, line 21 -page 22, line 29 --- -/--	1-28

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents:

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L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

O document referring to an oral disclosure, use, exhibition or other means

P document published prior to the International filing date but later than the priority date claimed

T later document published after the International filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

X document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

& document member of the same patent family

Date of the actual completion of the International search

20 September 2001

Date of mailing of the International search report

27/09/2001

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/IB 01/01068

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>WO 01 34815 A (CAMBRIA BIOSCIENCES, LLC) 17 May 2001 (2001-05-17) page 5, last paragraph -page 6, paragraph 4 page 20, paragraph 2 page 24, last paragraph; example 1 page 13, last paragraph -page 15, paragraph 2</p> <p>-----</p>	<p>1-10,12, 22,23,25</p>

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/IB 01/01068

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			EP 1104489 A1	06-06-2001
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			WO 0001846 A2	13-01-2000
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			NO 20010019 A	05-03-2001
WO 0134815	A	17-05-2001	AU 1461701 A	06-06-2001
			WO 0134815 A1	17-05-2001